

REVIEW

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The cGAS–STING pathway: more than fighting against viruses and cancer

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Abstract

In the classic Cyclic guanosine monophosphate–adenosine monophosphate (cGAMP) synthase (cGAS)–stimulator of interferon genes (STING) pathway, downstream signals can control the production of type I interferon and nuclear factor kappa-light-chain-enhancer of activated B cells to promote the activation of pro-inflammatory molecules, which are mainly induced during antiviral responses. However, with progress in this area of research, studies focused on autoimmune diseases and chronic inflammatory conditions that may be relevant to cGAS–STING pathways have been conducted. This review mainly highlights the functions of the cGAS–STING pathway in chronic inflammatory diseases. Importantly, the cGAS–STING pathway has a major impact on lipid metabolism. Different research groups have confirmed that the cGAS–STING pathway plays an important role in the chronic inflammatory status in various organs. However, this pathway has not been studied in depth in diabetes and diabetes-related complications. Current research on the cGAS–STING pathway has shown that the targeted therapy of diseases that may be caused by inflammation via the cGAS–STING pathway has promising outcomes.

Keywords: Anti-virus inflammation, cGAS–STING pathway, Diabetes, Insulin resistance, Diabetes complications

Introduction

In the past 20 years, metabolic syndrome (MetS) has evolved from a lesser understood term to a commonplace topic. The MetS has also become a risk factor for many chronic diseases [1]. Evidence confirms that the MetS and inflammation are closely related [1–3], and the link between the MetS and inflammation has provided very interesting ideas about the treatment of the MetS. As for diabetes mellitus (DM), a representative type of MetS, current studies also demonstrate that although the etiology of diabetes is not fully clear, inflammation and autoimmunity contribute significantly to the development of diabetes [4–6], and treatment of the inflammatory response to diabetes has shown beneficial therapeutic effects [7]; therefore, further research on the pathogenesis of diabetes and its complications is necessary.

Inflammatory responses and impaired mitochondrial function have been implicated in the function of many organs, and many of these effects are detrimental. Recently, investigations on the cyclic guanosine monophosphate–adenosine monophosphate (cGAMP) synthase (cGAS)–stimulator of interferon genes (STING) pathway have increased with a focus on its association with innate immunity. Initially, the cGAS–STING pathway was primarily concerned with immune mechanisms in infectious diseases [8, 9]. As research progressed, researchers identified impressive expressions of the cGAS–STING pathway in cancer and autoimmune diseases as well [10–12]. Recently, surprisingly, some work has demonstrated that the cGAS–STING pathway is highly associated with lipid metabolism [13, 14]. Therefore, an investigation of whether the cGAS–STING pathway is participating in and influencing the course of diabetes and its subsequent complications is warranted. Here, we will selectively summarize some literature reports on the effects of the cGAS–STING pathway in different organ diseases and focus on the

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possible cross-talk between the cGAS–STING pathway and diabetes. Since there is no clearer direct evidence of the link between the cGAS–STING pathway and diabetes, through some indirect literature reports, we can still observe an inextricable link between the cGAS–STING pathway and diabetes and its complications. This review may provide useful insights into the precise treatment of diabetes and may help guide future research.

The cGAS–STING pathway

STING (transmembrane protein 173, TMEM173 and MPYS/MITA/ERIS) is an evolved consensus transmembrane protein found in the endoplasmic reticulum (ER) of mammals and protozoa [15]. When STING was first discovered in 2008, researchers were investigating open reading frames that induce antiviral gene expression [16, 17]. Coincidentally, some researchers discovered that STING is involved in the transduction of proapoptotic signals [18]. Subsequently, STING was reported to exert antiviral effects through the TANK-binding kinase 1 (TBK1)-interferon regulatory factor 3 (IRF3) axis [16, 17], but the proteins related to its upstream signaling function were not confirmed at that time. Later, discoveries led by Chen confirmed that cGAS is a direct DNA receptor. Upon direct binding of cytoplasmic DNA, cGAS forms a dimer that catalyzes the production of 2′/3′-cGAMP from ATP and GTP, and then the binding of cGAMP activates STING, which facilitates the mobilization of downstream molecules [19, 20].

Recent works have also identified several other critical functions of the cGAS–STING pathway, which may also be involved in antiviral mechanisms and other chronic disease responses. For example, the cGAS–STING pathway was demonstrated to have a role in cancer and lipid metabolism as well as in a range of chronic inflammation-related diseases in various organs [10, 13, 14, 21–23]. Ongoing investigations of the cGAS–STING pathway suggest that agonists or inhibitors associated with this pathway are effective in the treatment of several inflammation-induced diseases.

The activation of the cGAS–STING pathway and its features

cGAS, a cytoplasmic DNA sensor or receptor that binds directly to DNA, is composed of approximately 520 amino acids [19]. It has a stable and active dimeric structure and its interactions with DNA have been confirmed in X-ray crystallographic studies [26, 27]. Dimer formation is an allosteric process and changes the state of cGAS from inactive to active [28, 29]. Notably, double-stranded DNA (dsDNA) can be sandwiched between these dimers [19].

When cGAS detects dsDNA from various sources (e.g., viral and bacterial DNA [19], mitochondrial DNA (mtDNA) [30], and micronuclear DNA [31–34], etc.), as a response, it is activated and catalyzes the formation of 2′/3′-cGAMP through a series of catalytic reactions and conformational changes (Fig. 1). These reactions are simply divided into two steps: in the first step, cGAS uses adenosine triphosphate (ATP) and guanosine triphosphate (GTP) as precursors and moves ATP to the 2′OH of GTP to generate linear isodinucleotide phosphate pppG (2′–5′) pA, which serves as the base material for the second step [35]; cGAS transfers the GTP to the 3′OH of adenosine phosphate to finally generate cGAMP [36]. The activation of cGAS is not modulated at the transcriptional level, but is modified at the post-translational level, while a number of other mechanisms are involved, including through ubiquitination, acetylation, phosphorylation, and cleavage by cysteinase. These modifications regulate cGAS activity by different mechanisms, e.g. direct modification, removal of the active site, stabilization of the protein, etc. [37–48].

cGAS activates STING and its downstream pathways via cGAMP. It has been shown that inactive STING dimers are present in the ER, and many TBK1 molecules can bind to STING dimers to form STING-TBK1 inactive complexes [49, 50]. Upon binding of cGAMP to the STING-TBK1 complex, the STING

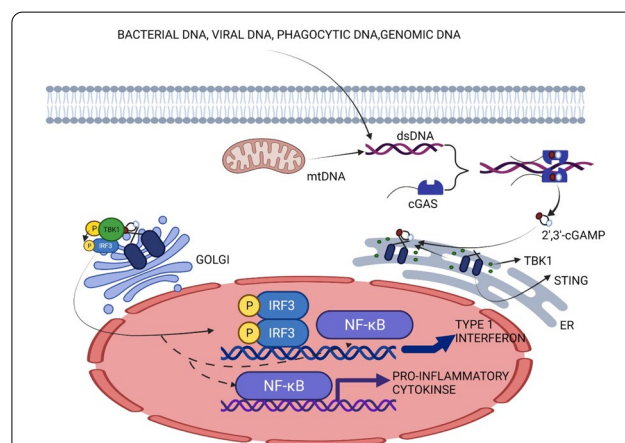


Fig. 1 Activation of the cGAS–STING pathway in cells. cGAS is stimulated by foreign double-stranded DNA, mtDNA in the cytoplasm, and phagocytosed DNA. Occupation of the substrates ATP and GTP catalyzes the formation of cGAMP via the induction of cGAS, which subsequently binds to STING located on the ER membrane. The incorporation of cGAMP into STING facilitates the trafficking of STING to the Golgi apparatus. Over the course of translocation, STING recruits and activates TBK1, which subsequently catalyzes the phosphorylation and nuclear translocation of IRF3 and, to a lesser extent, of NF-κB, resulting in the increased synthesis of IFN, along with other inflammatory molecules

dimer undergoes a spatial transition and triggers phosphorylation/activation of the transcription factor IRF3 [49, 50] (Fig. 1). Once IRF3 is activated by the active STING-TBK1 complex, it enters the nucleus and activates downstream reactions [51, 52]. Of note, STING can induce nuclear factor kappa-light chain-activated B-cell enhancer (NF- κ B), but in experiments it was shown that the human and mouse STING alleles induced a strong IRF3 response and a relatively weak NF- κ B response [53].

Downstream signaling of the cGAS–STING pathway controls interferon (IFN) and NF- κ B (a transcription factor that drives expression of pro-inflammatory genes including cytokines and chemokines) expression and promotes the activation of proinflammatory molecules [53–55]. Activated IFN can lead to the upregulation of several hundred IFN-stimulated genes (ISG) [56–58], which in turn promotes the secretion of several pro-inflammatory cytokines. These substances can limit viral replication and induce apoptosis.

Several studies have revealed a promising potential of cGAS use in non-infectious diseases. For instance, in addition to catalyzing the production of second messengers in the nucleus, cGAS can also restrain the restoration of DNA double-strand breaks (DSBs) through homologous recombination without STING or the catalytic activity of cGAS [37, 38]. This is supported by results indicating that the knockdown of cGAS inhibits both DNA damage and tumor outgrowth in vitro and in vivo [37, 38]. Preliminary studies have also shown that only adequately long dsDNA can make the cGAS oligomers bind to DNA with high affinity and produce subsequent activity [37]. Based on the above findings, cGAS can be considered as an antitumor factor, while the targeted recruitment of cGAS can be a potential new direction for tumor therapy.

STING can initiate autophagy [59, 60], and it promotes cell survival by maintaining normal autophagic activity after hydrogen peroxide treatment [61]. Despite the fact that STING is implicated in the mediation of autophagy, there is a lack of accurate insight into the mechanism of STING-induced autophagy. Similarly, the cGAS–STING pathway participates in the regulation of cellular senescence [33, 62], an irrecoverable cell cycle stalling that occurs in response to severe cellular stress. In addition to inhibiting auto-proliferation, senescent cells produce inflammatory cytokines, growth factors, and proteases. This phenotype is known as the senescence-associated secretory phenotype (SASP); it strengthens senescence in an autocrine mechanism and mediates growth inhibition via a paracrine mechanism. In addition, chemokines of SASP enable the activation and recruitment of immune cells to destroy abnormal cells with damaged DNA [63,

64]. In the absence of cGAS as well as STING, both SASP induction and immune cell infiltration are defective [62].

Finally, the cGAS–STING pathway can cell type-specific induce apoptosis [65], STING can induce the expression of many pro-apoptotic molecules [66]. In the cGAS–STING pathway, activated IRF3 can combine with BCL-2 related X protein (BAX) to form a complex that directly induces apoptosis [67]. Furthermore, STING is also involved in the induction of the lytic cell death program [68]. It can activate mixed-lineage kinase domain-like pseudokinase (MLKL) [69], an enzyme involved in the induction of necrosis. p53 upregulated modulator of apoptosis (PUMA) [70] is activated in an MLKL-dependent manner, which exacerbates mtDNA leakage in mitochondria and thus leads to programmed cell death.

Major identified diseases associated with the cGAS–STING pathway

The cGAS–STING pathway and infectious diseases

The cGAS–STING pathway is primarily activated by dsDNA that enters the cytoplasm abnormally; therefore, it is tempting to emphasize the connection between the cGAS–STING pathway and viral infections, such as those caused by classical herpes simplex virus type 1 [71]. Considerable research results are available, but as they have little relevance to the topics described in this review, only some reviews are provided for reference only [8, 9]. Notably the study found that mouse hepatocytes lack STING, but the application of selective expression of STING in mouse hepatocytes elevated the IFN response to DNA stimulation and enhanced the control of hepatitis B virus (HBV) [72].

The cGAS–STING pathway and cancer

Given how sensitive the cGAS–STING pathway is to abnormal intracellular DNA, researchers are interested in investigating the effects of this pathway on cancer. As research has progressed, scientists have obtained fruitful results regarding the associations between the cGAS–STING pathway and tumors. Given that it is not the subject of this review, here, the authors provide some reviews on the relationship between the cGAS–STING pathway and tumors [10, 11]. Among the studies on the relationship between the cGAS–STING pathway and tumors, it was surprising that researchers found that the cGAS–STING pathway might be relevant to some tumor metastasis [73]. The discovery reflects the complexity of the cGAS–STING pathway in regulating disease processes and alerts researchers to the necessity of more rational application of cGAS–STING pathway-related drugs for treatment.

The cGAS–STING pathway and autoimmune diseases

Autoimmune diseases are conditions that arise when the body abnormally attacks its own normal cells. At least 80 such disorders have been identified, and they can occur in almost any part of the body. Common symptoms include intermittent fever and fatigue. Some autoimmune diseases have been linked to mutations in genes that regulate the cGAS–STING pathway. For example, patients with vascular and pulmonary syndrome (VAPS) [74], a systemic inflammatory disease, have mutations in STING exons. Other genetically induced autoimmune diseases include STING-associated vasculopathy with onset in infancy (SAVI) [74–77]; Aicardi-Goutieres syndrome [78, 79], systemic lupus erythematosus (SLE) [80, 81], familial cerebellar lupus erythematosus [82, 83], and polyarthritis induced by deoxyribonuclease II (DNaseII) gene defects [84–86]. Table 1 lists these diseases and the genes involved.

Because early knowledge of the cGAS–STING pathway was not comprehensive initially, the studies conducted mostly focused on viral diseases, cancer, and autoimmune diseases. However, as research on the cGAS–STING pathway progressed, researchers tended to link the cGAS–STING pathway to chronic diseases. Although the cGAS–STING pathway is still most intensively studied in relation to viruses, targeting the cGAS–STING pathway in exploring the pathogenesis of chronic diseases is an attractive research direction.

Novel physiological and pathological roles of the cGAS–STING pathway in specific organs

With the knowledge of mechanisms related to the cGAS–STING pathway, researchers have gained a better understanding of diseases caused by low-grade inflammation, and as a result, some investigators have initiated investigations on the cGAS–STING pathway in different

organs. Some of the research advances in these areas are presented below.

Heart

Heart disease is one of the most common causes of death worldwide, and the etiology of many common heart diseases is still not fully understood, but inflammation, fibrosis, and oxidative stress are known to contribute to heart-related diseases. Given the impressive performance of the cGAS–STING pathway in inflammation, many research groups have directed their work to the function that the cGAS–STING pathway performs in different cardiac diseases.

First identified was that the cGAS–STING pathway was more expressed in the hearts of SLE patients compared to normal subjects [87]. In a subsequent study, a research group found that STING-IRF3 was activated and induced an inflammatory response in endothelial cells treated with palmitic acid (PA) [88]. In addition to the cGAS–STING pathway being shown to be activated in vitro, the cGAS–STING pathway was shown to exacerbate the pro-inflammatory response in a mouse model of myocardial infarction [89]. In contrast, myocardial function and survival were improved in myocardial infarct mice when cGAS and STING were absent [89]. Not only limited to myocardial infarction, high expression of the cGAS–STING pathway in different causes of cardiac dysfunction has been shown to be involved in the development of cardiac dysfunction, and cardiac function is significantly protected when the cGAS–STING pathway can be inhibited [90–92].

Not only limited to cardiac energy metabolism and the coronary system, the cGAS–STING pathway also contributes to the abnormal cardiac function caused by structural abnormalities of the heart. STING was found to be more highly expressed in cases of dilated cardiomyopathy (DCM) and hypertrophic cardiomyopathy (HCM) as well, and cardiac hypertrophy and fibrosis were significantly attenuated in DCM and HCM when STING was inhibited. This result suggests that STING serves as a pivotal link in pathological myocardial hypertrophy [93]. The effect of the cGAS–STING pathway on myocardial structural changes is not limited to cardiomyopathy but is also seen in transverse aortic systolic surgery (e.g., mouse models) [23]. In contrast, when cGAS expression is suppressed, fibrosis, myocardial hypertrophy and apoptosis are reduced, and cardiac function is significantly improved [23].

Researchers have found some clues as to the mechanism by which the cGAS–STING pathway functions in the heart, but further studies are still needed to clarify the mechanism. For instance, leakage of mtDNA into cytoplasm is thought to be a possible factor in the activation

Table 1 cGAS–STING-related autoimmune diseases

STING-related gene mutations	TREX1 mutation	DNase II mutation
VAPS	AGS	Polyarthritis
SAVI	SLE	
	Familial cerebellar lupus erythematosus	
	Retina with leukodystrophy	Vascular disease

VAPS vascular and pulmonary syndrome, AGS Aicardi-Goutieres syndrome, DNaseII deoxyribonuclease II, SLE systemic lupus erythematosus, SAVI STING-associated vasculopathy with onset in infancy, TREX1 three prime repair exonuclease 1

of the cGAS–STING pathway in the heart. Studies have shown a clear association between mtDNA and cardiac dysfunction [94]. Whereas in earlier studies mtDNA stress was found to trigger a significant antiviral innate immune response, mtDNA leakage in the cytoplasm could enhance the cGAS–STING pathway and thus the inflammatory response [30]. In studies on endothelial cells, researchers found that PA activates the cGAS–STING pathway by causing mitochondrial damage and mtDNA release into the cytoplasm [88]. This study did not further confirm the link between mtDNA and cGAS–STING pathway in animal models, but the results of this experiment also provide ideas for the mechanism of cGAS–STING pathway activation in heart disease. ER stress has also been suggested as a crucial mechanism for the role of the cGAS–STING pathway in cardiac diseases. Cardiac hypertrophy and fibrosis were significantly attenuated when STING was inhibited in mice with cardiac hypertrophy [93], whereas there was a significant increase in STING expression when ER stress activators were applied in *in vitro* experiments [93]. These findings suggest that alterations in STING expression correlate significantly with ER stress and may promote the progression of cardiac hypertrophy when STING expression is increased. Therefore, it is necessary to further investigate the link between the cGAS–STING pathway and ER stress and its mechanisms in appropriate models.

In addition to mtDNA leakage and ER stress, researchers found in mouse models of myocardial infarction that the expression of some key inflammatory substances (e.g., inducible nitric oxide synthase; iNOS) was reduced in the absence of cGAS and STING and could contribute to the conversion of macrophages to a repair phenotype, which improved the survival of myocardial infarction mice [25, 95]. These experimental results suggest that the cGAS–STING pathway may be involved in cardiac disease by altering macrophage polarity.

In **conclusion**, the cGAS–STING pathway exhibits a function in cardiac disease as exacerbating cardiac disease, and the mechanisms by which the cGAS–STING pathway is involved in cardiac disease are complex, requiring researchers to conduct additional studies to elucidate the role played by the cGAS–STING pathway in the progression of cardiac disease. This may also provide important assistance for the precise use of inhibitors related to the cGAS–STING pathway.

Lung

Many lung diseases are thought to be correlated with inflammation and autoimmune hyperactivation, and the cGAS–STING pathway has been introduced into the exploration of the mechanisms of these lung diseases because of its manifestation in diseases associated with

inflammation. These diseases include chronic obstructive pulmonary disease (COPD), pulmonary fibrosis and asthma.

COPD is an aggressive, chronic inflammatory disease related to the impairment of lung function, which is characterized by persistent long-term inflammation, typically as a result of excessive production of reactive oxygen species, leading to oxidative stress and DNA damage. There have been some developments regarding the association between COPD and the cGAS–STING pathway. In one study, researchers found that cigarette smoke caused respiratory barrier damage in mouse model, which resulted in the release of DNA into the cytoplasm. This activated the cGAS–STING pathway, which in turn triggered type I IFN-dependent lung inflammation. These effects were significantly attenuated in mice lacking STING [97].

However, another group of researchers found that STING expression was decreased in mice exposed to cigarette smoke or smoke extracts. The experiments were conducted by exposing mice to cigarette smoke to develop a COPD model and then infecting the mice with recombinant adenoviral vectors (rAdVs) to develop a model of acutely exacerbated COPD (AECOPD). In the experiment, STING expression was suppressed in mice exposed to cigarette smoke + infected with rAdVs compared to rAdVs-infected mice. In addition when STING was knocked out, the lung inflammatory response and fibrosis were increased in cigarette smoke + rAdVs infected treated mice [98]. This result suggests that STING can protect lung function in rAdVs-infected mice under cigarette smoke exposure. Not limited to animal models, another study confirmed that ISG expression was detectable in both stable COPD patients and healthy subjects, while ISG expression was reduced in patients with AECOPD [96]. Based on the above experimental results, we conjecture that, given the role of STING in both antiviral and inhibition of pulmonary fibrosis, STING is used to fight viral infection when the body is infected with a virus, and the corresponding inhibition of inflammation and fibrosis may be compromised, which may lead to an exacerbation of disease progression in AECOPD. The mechanism for the different results in the two groups of mice exposed to cigarette smoke is also unclear and may be related to the different duration of smoke treatment in the two groups. What is confirmed is that STING can have a relatively important contribution to disease progression in COPD, and that it may not play the same role in acute and chronic COPD.

In addition to its involvement in COPD, cGAS–STING also plays a significant role in pulmonary fibrosis. Silica particles cause lung inflammation and fibrosis, and in a 2018 study, STING was experimentally confirmed as a critical component in silica-induced

lung disease. Patients with silicosis had significantly higher plasma levels of dsDNA than normal subjects, and the cGAS–STING pathway was activated in the lungs of these patients and mice exposed to silica particles [99]. In animal experiments, silica particles triggered lung cell death and the release of dsDNA into the bronchoalveolar space leading to STING expression, which may be a possible mechanism by which STING is activated [99]. Idiopathic pulmonary fibrosis is a fatal interstitial lung disease, a form of pulmonary fibrosis characterized by the irreversible destruction of lung structures, which in turn leads to respiratory failure and death. A group of investigators found that ER stress was triggered, and STING expression was decreased in patients diagnosed as having acute exacerbations of idiopathic fibrosis and in pulmonary fibrosis mouse models. When treated, STING levels significantly increased in patients with pulmonary fibrosis [100]. Reportedly, STING deficiency leads to increased collagen deposition in the lung and increased pulmonary fibrosis, reflecting the protective role of STING in idiopathic pulmonary fibrosis [101]. The different expression levels of STING in silica-induced and idiopathic pulmonary fibrosis show that there may be different cGAS–STING regulatory mechanisms for different etiological conditions. This requires further investigation of the various regulatory mechanisms involved.

Asthma is a chronic inflammatory lung disease involving multiple cells (e.g., eosinophils, mast cells, T lymphocytes, neutrophils, and airway epithelial cells) and cellular components. It is characterized by airway hyperresponsiveness and reversible impaired ventilation. Common exacerbation triggers include viral infections and dust mites. In an experiment conducted in an asthma mouse model, the accumulation of cytoplasmic dsDNA in airway endothelial cells increased in mice in an allergic state. Furthermore, the absence of cGAS significantly attenuated allergic airway inflammation caused by chicken ovalbumin or house dust mites (HDM) [102]. Therefore, the cGAS–STING pathway probably contributes to the immune response at the onset of asthma; However, whether cGAS itself is involved in this immune response, or whether cGAS activates the immune response through the cGAS–STING pathway, further studies are necessary to confirm. Other groups have also examined gene expression in patients with mild and severe asthma and confirmed that ISG levels were increased in these patients and that increased airway ISG expression was associated with decreased lung function. More importantly, by detecting ER stress-related proteins, the experiments show that ISG was associated with ER stress markers through the detection of ER stress-related proteins, implying that there may be a potentially

link between ISG-induced inflammation and ER stress [103], which requires further de-confirmation.

In conclusion, further confirmation of the function of the cGAS–STING pathway during the development of various lung conditions is required. It is also crucial to further elucidate the effect of cGAS–STING pathway-mediated cytokine secretion in patients with pulmonary conditions.

Liver

Numerous studies have confirmed the link between the liver and the cGAS–STING pathway. In addition to HBV and liver cancer, the cGAS–STING pathway has been shown to be activated in other kinds of liver diseases (e.g., non-alcoholic fatty liver disease (NAFLD), alcohol liver disease (ALD), liver ischemia–reperfusion, etc.).

In the previous section, we have described that STING-IRF3 can be activated in PA-induced endothelial cells, and this experiment also reported that in high-fat diet (HFD) mice, STING-IRF3 is also activated and that both HFD-induced inflammation and obesity are suppressed to some extent when STING is absent [88]. And in different models of nonalcoholic steatohepatitis, STING deficiency attenuated hepatic steatosis, inflammation and fibrosis [104]. In addition to animal models, it has also been demonstrated that increased STING expression has been observed in Kupfer cells from nonalcoholic steatohepatitis patients [105]. A report of a trial on 98 participants confirmed that patients with NAFLD had higher levels of STING in liver tissue, along with higher levels of inflammation and fibrosis, compared to the control group [106].

ALD is a common indication for liver transplantation. In a 2016 study on liver fibrosis in mice, ER stress promoted IRF3 activation and resulted in hepatocyte apoptosis and injury. Conversely, both fibrosis and apoptosis were inhibited in mice lacking IRF3 expression due to knocking out STING or IRF3 [107]. This confirmed the role of STING in promoting the progression of liver disease to fibrosis or hepatocyte death in mice. In subsequent studies on ALD, investigators established that the cGAS–STING pathway gets occupied and leads to IRF3 interaction with proapoptotic BAX, which resulted in hepatocyte apoptosis, thus setting the stage for inflammation and injury [108]. Moreover, liver damage was reduced when the cGAS–STING pathway was inhibited [109]. Alcohol-induced cytoplasmic leakage of mtDNA amplifies the cGAS–STING pathway, which has been shown to amplify liver injury through hepatic gap junctions in ALD [109]. In view of the above experimental results, although the mechanism of the effect of the cGAS–STING pathway on fatty liver (including NAFLD and ALD) is not fully elucidated, it is possible to suggest

that the cGAS–STING pathway could be a potential therapeutic target for the treatment of fatty liver.

With the development of liver transplantation, liver ischemia–reperfusion has received increasing attention. Of note, individual proteins of the cGAS–STING pathway are implicated in this process. The researchers found that cGAS itself can have a protective effect on the liver. This protective property is independent of STING, a process that protects hepatocytes from apoptosis after oxidative stress [110]. Another study in 2020 with a mouse model of ischemia–reperfusion confirmed that in macrophages, STING can promote the activation of NLRP3 which can exacerbate liver ischemia–reperfusion injury [111]. Taken together, the cGAS–STING pathway is of great influence on the occurrence of diseases in the liver. These studies on cGAS–STING in the liver provide new insights into our understanding of the largest visceral organ.

Kidneys

Acute or chronic kidney lesions may be associated with the cGAS–STING pathway. As of 2017, a study conducted on SLE by a group of researchers confirmed that tumor necrosis factor (TNF)-like weak inducer of apoptosis could activate type I IFN in lupus nephritis (LN) causing kidney injury. This demonstrated that type I IFN has a role in kidney injury caused by autoimmune diseases [112]. In a subsequent study, Japanese scholars found that in cisplatin-induced acute kidney injury mouse model, the cGAS–STING pathway got triggered by mtDNA and elicited inflammatory injury [113]. Conversely, the suppression of inflammation was observed in *STING*-knockout mice, thus confirming the usefulness of the cGAS–STING pathway in the pathogenesis of acute kidney disease [113].

In a 2019 study, researchers also affirmed the contribution of the cGAS–STING pathway to the development of chronic kidney disease. Researchers initially observed that chronic kidney disease recruits activation of the cGAS–STING pathway, and that this activation is associated with abnormalities in mitochondrial metabolism [114]. Subsequently, they also observed that fibrosis and renal failure were suppressed in *STING* knockout mice [114]. These experiments confirm the critical role of the cGAS–STING pathway in renal inflammation due to abnormal mtDNA accumulation, but do not provide sufficient evidence to define whether the mobilization of the cGAS–STING pathway is dependent on the amount of mtDNA leakage. Another study on the development of LN-related end-stage renal disease in African-Americans also confirmed the essential role of the cGAS–STING pathway in renal disease [115]. This pathway involved IFNs plus nucleosome-associated dsDNA (nsDNA)

fragments and triggered the expression of IFN- β . Furthermore, the inhibition of STING suppressed the over-expression of human nsDNA.

In summary, the abovementioned findings demonstrate the vital role the cGAS–STING pathway play in acute to chronic kidney lesions by increasing the expression of individual cGAS–STING pathway-associated proteins upon the occurrence as well as suppression of damage in models after cGAS–STING pathway-associated proteins are knocked out. This indicates that precise targets of the cGAS–STING pathway can be potentially useful in kidney-related diseases.

Other organs

There is now some evidence that the cGAS–STING pathway is activated in diseases in organs other than those discussed above; for example, in the nervous system, the cGAS–STING pathway is activated in traumatic brain injury, ischemic stroke, and Huntington's disease [116–118]. Moreover, the activation of the cGAS–STING pathway is also correlated with diseases such as age-related macular degeneration, aortic degeneration, small angiogenesis, acute pancreatitis, and colitis [119–123].

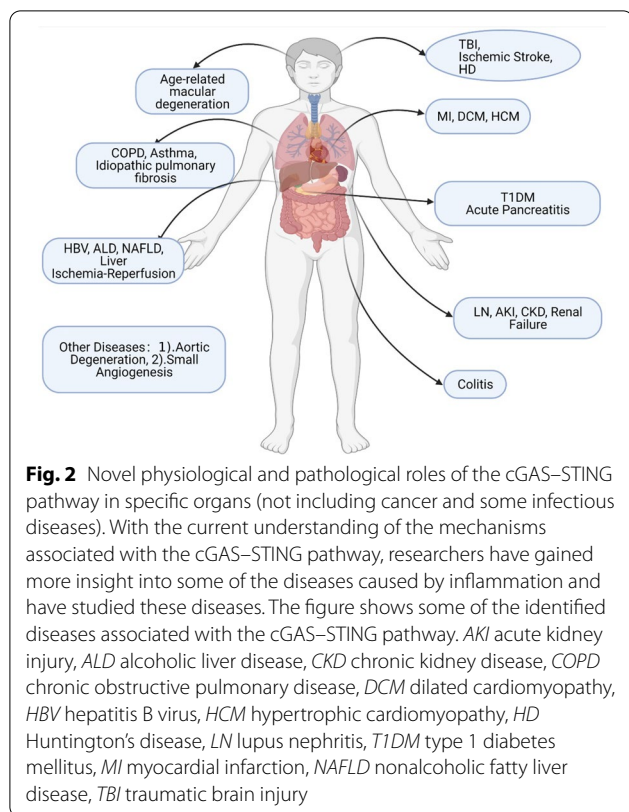
Overall, the cGAS–STING pathway exerts a complex impact on the progression of these diseases, not only via inflammatory damage promotion, but also, in some respects, through a protective function, making it difficult to identify the pathway that is more important in the development of the diseases. We can speculate that the inhibition of cGAS–STING in multiple organs (Fig. 2) may reduce the severity of some diseases; however, it may also significantly downregulate innate immune function, leading to an increased risk of viral infection and tumor invasion. Therefore, in addition to investigating the mechanisms of action of cGAS–STING, it is important to study how the body precisely regulates the pathways involving these proteins.

Role of the cGAS–STING pathway in diabetes

Diabetes imposes a significant economic burden on individuals and the society. This is attributed to the increased prevalence of diabetes and the increased cost per patient with diabetes [124]. Although there have been tremendous advances in diabetes research in recent years, including extensive research on the mechanisms and treatment of diabetes, a high prevalence of diabetes and related complications remains. Therefore, there is an urgent need to develop more effective therapies.

Diabetes and inflammatory responses

diabetes is a metabolic disease affecting multiple organs and is mainly caused by absolute or relative insulin deficiency which mainly manifests as reduced glucose



tolerance and hyperglycemia. Empirical evidence indicates that inflammatory pathways can contribute to obesity, insulin resistance (IR), and its subsequent diabetes-related metabolic disorders and complications. Therefore, the targeted administration of anti-inflammatory therapy may be helpful in the prevention of diabetes, and the application of tests to detect relevant inflammatory indicators may help understand the prognosis of diabetes and metabolic disorders.

Type 1 diabetes mellitus (T1DM) is an autoimmune disease characterized by the destruction of pancreatic islet β -cells leading to inadequate insulin synthesis. The involvement of immune cells in the progression of T1DM has been demonstrated in several animal models. As in the case of the T1DM model of transmissible T cell transfer, it has been shown that T cells are capable of destructive peri-islet inflammatory infiltration and diabetes induction [125, 126]. In addition, macrophages are key mediators of islet inflammation owing to their ability to secrete cytokines such as IL-1 β and TNF- α . These two cytokines can act synergistically with IFN- γ , leading to the uptake of iNOS and the consequent generation of nitric oxide (NO) [127]. The mechanisms mentioned above may lead to pancreatic β -cell death in multiple ways; therefore, the control and regulation of local inflammatory cytokine production

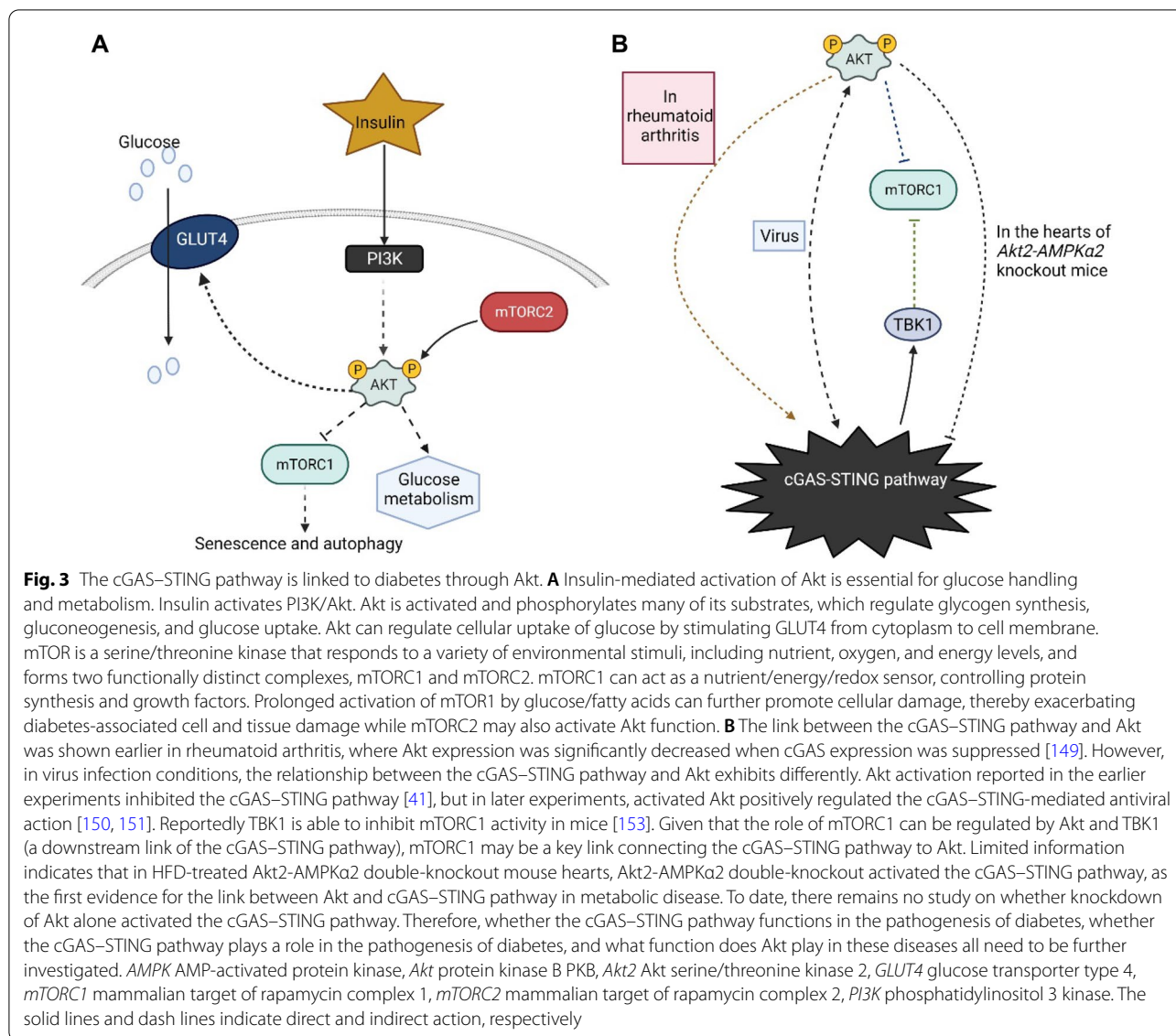
may be a key factor in determining the outcome of T1DM progression.

Type 2 diabetes mellitus (T2DM), in contrast, is characterized by a relative scarcity of insulin, which manifests as IR, a state in which essential organs, such as the liver and muscles, are less sensitive to insulin. Fat accumulation in the liver and muscle tissues is the antecedent of IR and precedes the onset of T2DM. Moreover, fat accumulation exacerbates IR and causes dysfunction of pancreatic β -cells at the late stage of T2DM [128]. It is now well-established that obesity and its associated MetS are positively associated with the induction of inflammatory responses (mediated by NF- κ B) which contribute to the development of IR and T2DM [129]. Adipokines (such as leptin and adiponectin) stimulate other inflammatory responses in individuals with obesity, and macrophages and immune cells can then infiltrate adipose tissue [130–133]. A combination of these factors results in low levels of sustained inflammatory responses in the body, eventually leading to diseases related to organ damage and dysfunction.

Therefore, it is imperative to focus on diabetes prevention and treatment research. Notably, inflammatory pathways are under consideration as treatment targets in diabetes and its complications.

Protein kinase B (Akt) and diabetes

Insulin is the most important substance in metabolism regulation because it regulates the production of glucose, lipids, and proteins mainly through the phosphatidylinositol 3-kinase (PI3-K) pathway [134]. Substrate proteins of the insulin receptor are phosphorylated, resulting in binding and activation of PI3-K, which activates downstream kinases, like phosphatidylinositol-dependent kinases [134], and consequently initiates Akt expression (Fig. 3). Akt is an important protein involved in insulin signaling, metabolism, cell growth, and the cell cycle. It mediates glucose transportation via glucose transporter-4 (GLUT-4) molecules into cells that metabolize glucose in an insulin-dependent manner [134]. Akt1 inhibits apoptosis and improves cell survival [135]. Akt2 is involved in the insulin signaling pathway [136] as evidenced by the development of IR and T2DM-like symptoms in human with an Akt2 deficiency [137]. Although there are reviews that describe the role of Akt in diabetes [138], additional studies now suggest that the regulation of Akt in diabetes may not be as simple a process as considered previously. mTOR is a serine/threonine kinase that responds to various environmental stimuli, including nutrients, oxygen, and energy levels and forms two functionally distinct complexes, mTORC1 and mTORC2. The mTOR pathway integrates inputs



from upstream pathways, including insulin, growth factors and amino acids, while mTOR also senses cellular nutrient, oxygen and energy levels. mTORC1 can act as a nutrient/energy/redox sensor that controls protein synthesis and growth factors that can trigger the activation of mTORC2, whereas mTORC2 can activate Akt1 which can activate mTORC1 indirectly (Fig. 3) [139]. The chronic activation of mTOR by glucose/fatty acids can further promote the apoptosis of pancreatic β-cells, thereby exacerbating diabetes progression. From the above brief description, we can find that the function of the Akt protein family is very complex and may contain multiple mechanisms; and shows crosstalk with many other signaling pathways. These complex mechanisms are focused in diabetes as shown by the remarkable

contribution that the Akt protein family can make in suppressing IR and increasing insulin sensitivity.

The complex functions exhibited by the Akt protein family undoubtedly endow the cells with powerful capabilities. However, this complex crosstalk also makes it difficult to predict the possible network-wide effects of diabetes treatment targeting pathway nodes. Given the diverse roles of the Akt protein family in diabetes, we must continue to investigate and uncover all possible feedback mechanisms and pathway interactions. It is equally essential to understand the initiating factors of diabetes and how these factors impact these mechanisms and interactions as well as how the various Akt-interacting signaling pathways support and compensate for each other.

Adiponectin and disulfide bond A oxidoreductase-like protein (DsbA-L)

Adipocytes secrete various adipocytokines; adiponectin, in particular, is a protein that plays a key role in sugar and lipid metabolism. It has 244 amino acids and has been identified as a crucial factor for the regulation of energy homeostasis through the employment of peroxisome proliferator-activated receptors (PPARs) [140] and AMP-activated protein kinase (AMPK) [141]. Owing to its key part in metabolism, adiponectin is considered an important therapeutic target in diabetes and MetS.

PPAR promotes the expression of adiponectin genes in diabetic mice and improves insulin sensitivity by increasing fatty acid utilization and reducing triglyceride levels in the liver and skeletal muscles [140, 142]. Additional evidence of decreased hepatic insulin sensitivity and poor response to agonists of PPARs in adiponectin-deficient mice suggests that adiponectin can slow the progression of diabetes by directly affecting insulin sensitivity [143]. In addition, the AMPK pathway can be activated by adiponectin to inhibit acetyl coenzyme A carboxylase, leading to increased fat oxidation and glucose transporter expression [144]. AMPK can promote glucose uptake by increasing GLUT-4 transport to the plasma membrane [145], whereas in adipose tissues, AMPK activation decreases PPAR expression and reduces adipogenesis by increasing lipolysis. It also suppresses the discharge of TNF- α and IL-6 in the adipose cells as well as increases the secretion of adiponectin [146]. In **conclusion**, adiponectin can mediate metabolic pathways in the body, and increasing its expression can reduce the risk of diabetes and MetS.

DsbA-L is a molecular chaperone involved in adiponectin multimerization and plays an important protective role in MetS [147, 148]. The adipose-specific overexpression of DsbA-L protects mice from HFD-induced adiponectin downregulation and reduces IR and hepatic steatosis. In addition to protecting mice from the adverse effects of obesity and IR by promoting adiponectin multimerization, DsbA-L appears to protect high-fat-fed mice through a adiponectin-independent pathway. In a study, mice with adiponectin knockout were divided into two groups, one overexpressing DsbA-L and the other serving as a control. No significant differences were observed in the body weights and insulin sensitivities of the same normal diet-fed mice. However, when both groups were fed an HFD, mice with DsbA-L overexpression and adiponectin knockout showed lower body weights and adipocyte volumes than mice with only adiponectin knockout [147]. In addition, the downregulation of DsbA-L expression reduces PPAR γ agonist function, suggesting that DsbA-L not only increases insulin sensitivity,

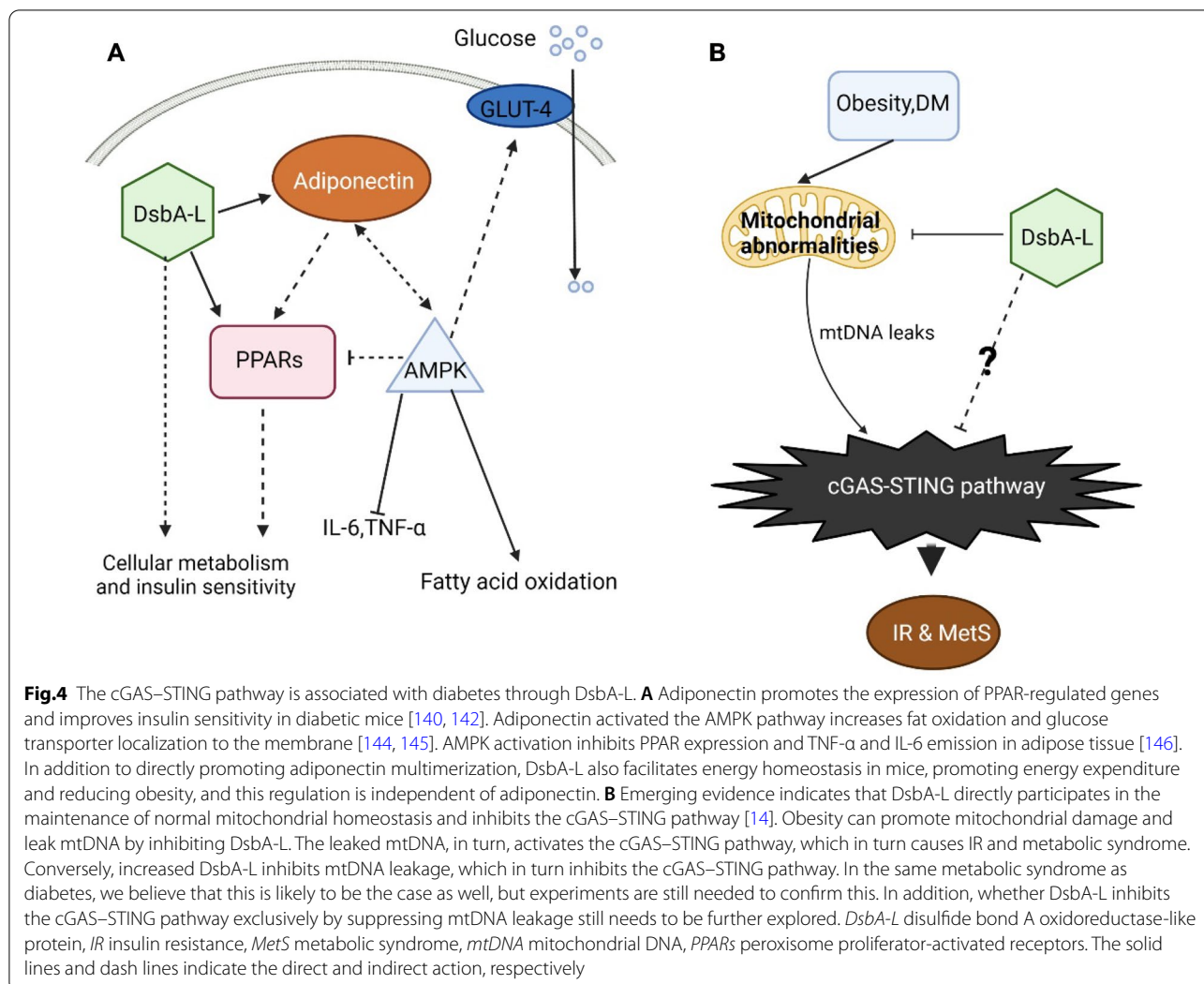
but also has a prominent function in the prevention of diabetes (Fig. 4) [148].

The protective function of adiponectin and DsbA-L in metabolism implies the essential use of adiponectin, DsbA-L, etc. as therapeutic targets when treating metabolism-related diseases such as obesity and diabetes. Although there are few studies on the relationship between DsbA-L and diabetes, it is necessary to investigate the molecular pathways through which DsbA-L inhibits the development of diabetes under the complex crosstalk of the whole network.

The cGAS–STING pathway is linked to diabetes through Akt

There is no direct evidence on whether the cGAS–STING pathway affects diabetes. The available evidence is based on animal experiments in non-obese diabetic (NOD) mice. Non-treated female NOD mice showed signs of T1DM from 16 weeks of age and the incidence of T1DM increased to approximately 90% at 35 weeks of age [24]; however, treatment of the female prediabetic NOD mice with DNA nanoparticles (DNPs) (activators of STING) at 4–8 weeks delayed the progression of T1DM, owing to which all mice remained disease-free until 27 weeks of age. Moreover, the prevalence of T1DM was also reduced at 35 weeks of age. However, this preventive effect was not sufficient if DNPs were administered at 8–12 weeks of age, indicating that it is possible to prolong the T1DM-free period and reduce the occurrence of T1DM by treating NOD female mice during early prediabetes [24]. Therefore, STING signaling may regulate autoimmunity in some way and thus reverse or prevent the progression and pathogenesis of T1DM. Since this report only addressed STING, further studies are warranted to determine the usefulness of the cGAS–STING pathway in diabetes. Moreover, since this study only focused on T1DM in NOD mice, it is urgent to address whether the cGAS–STING pathway can work in other types of diabetes.

The first experimental reports linking the cGAS–STING pathway to Akt was based on rheumatoid arthritis. Experimental results demonstrated that the cGAS–STING pathway promoted the inflammatory response in rheumatoid arthritis, and when cGAS expression was inhibited, Akt expression was significantly decreased (Fig. 3) [149]. Further research on the connection between the Akt and cGAS–STING pathways is needed. Given the unique advantages the cGAS–STING pathway provides in the antiviral mechanism, researchers are interested in the usefulness of the Akt pathway in the antiviral process of that pathway. The experimenters discovered that Akt has a negative effect in the cGAS-mediated antiviral immune response and that the activation of Akt resulted in the inhibition of cGAMP and



IFN- β production in the experiment [41]. This would appear to provide one aspect of the relationship between Akt and the cGAS–STING pathway. However, different experimental groups have confirmed from two different aspects that Akt would play a positive role in IFN-induced antiviral responses [150, 151]. Nevertheless, the above experiments all involve only the cellular level and do not fully explain the connection between Akt and cGAS–STING pathway, but it is safe to assume that there is a complex crosstalk between Akt and cGAS–STING pathway. In addition, Akt may be linked to the cGAS–STING pathway through mTOR, and this potential link between Akt and mTOR in energy metabolism has been described previously. There is now evidence that TBK1 can phosphorylate mTOR on S2159, thereby enhancing the production of IFN- β [152]. Furthermore, in *three-prime repair exonuclease 1* knockout mice, the activation of TBK1 inhibited mTORC1 activity [153]. Although

these studies have found an association between the Akt protein family and the cGAS–STING pathway in different diseases, none of them is direct evidence of a direct link between the Akt protein family and cGAS–STING in metabolic diseases.

Thus, when a study on HFD-induced cardiac abnormalities linked Akt2 to the cGAS–STING pathway, we gained a more direct understanding of the relationship between the Akt family and the cGAS–STING pathway [154]. The experiment investigated the cardiac deformation of Akt2 and AMPK associated with metabolic abnormalities via the double knockdown of *Akt2* and *AMPK α 2*. *Akt2-AMPK* double knockdown exhibited a strong potential for inducing autophagy and simultaneously increased the expression in the cGAS–STING pathway for HFD-fed mice, thereby exacerbating the inflammatory response in the heart and ultimately affecting cardiac function and structure (Fig. 3) [154]. In contrast, in

in vitro experiments, high levels of lipids increased the activation of the cGAS–STING pathway and promoted the inflammatory response of endothelial cells [89]. This study is the first to connect Akt proteins to the cGAS–STING pathway in metabolic diseases. However, since the study also involved AMPK α 2, further investigation is needed to confirm whether AMPK α 2 exists in the link between Akt2 and cGAS–STING. Given the protective ability of Akt2 in diabetes and the above mentioned studies, we tried to hypothesize that Akt2 can inhibit the expression of cGAS–STING pathway in diabetes progression and thus inhibit the inflammatory response induced by diabetes.

The cGAS–STING pathway is associated with diabetes through DsbA-L

Available evidence has demonstrated an association between DsbA-L and the cGAS–STING pathway in metabolic diseases. In 2017, Bai et al. [14] reported that the white adipose tissue (WAT) of HFD-induced obese mice showed significant elevation of cGAS and STING expression. This was attributed to the phosphorylation of TBK1, NF- κ B p65 and IRF3 and a marked increase in the expression of TNF- α in adipose tissue. The authors further demonstrated cGAS–STING pathway activation and increased mtDNA leakage and downregulation of DsbA-L expression. This suggests that obesity may downregulate mitochondrial DsbA-L expression and function, causing mitochondrial impairment and resulting in the mtDNA cytosol leak, activation of cGAS–STING pathway-mediated inflammatory responses, and eventually IR and MetS [14]. They further showed that *DsbA-L* knockdown alone, even in wild-type mice, compromises mitochondrial functionality and fosters the activation of the mtDNA-release-induced cGAS–STING pathway and the associated inflammatory responses. In contrast, DsbA-L overexpression protects against mtDNA leakage induced by obesity and consequently against the cGAS–STING signaling pathway (Fig. 4). When DsbA-L was deficient and when the cGAS–STING pathway was examined in adiponectin-specific knockout mice, no significant differences were found compared with the observations in DsbA-L-overexpressing adiponectin knockout mice. The exact underlying mechanism through which DsbA-L represses HFD-induced mitochondrial dysfunction and mtDNA liberation remained elusive. Furthermore, cGAS–STING signaling is enhanced and DsbA-L exposure is reduced in the WAT of diabetic (db/db) mice compared to that in control mice [14]. Overall, DsbA-L can inhibit the inflammatory response activated by the cGAS–STING pathway in MetS. However, whether the cGAS–STING pathway is directly correlated with DsbA-L needs to be further clarified, and whether this

inhibition is also present in diabetes still remains to be confirmed.

Potential effect of the cGAS–STING pathway in diabetes mediated through immune cells

T1DM occurs as a result of the T cell-mediated autoimmune disruption of pancreatic β cells, whereas T2DM is characterized by topical and systemic inflammation and facilitates the development of IR. In recent years, increasing evidence has shown that T cells are vital for the induction of metabolic inflammation and IR. A team of researchers, by way of example, reported the increase in the activated CD4 + T cell populations in the visceral adipose tissues of obese mice, which also show the expression of programmed death1 (receptor) and CD153, and exhibit cellular senescence [155]. The extensive evidence and the involvement of immune cells in the pathogenesis of diabetes and in its complications are well depicted in several reviews and are not discussed here [156–159]. However, to develop new therapeutic approaches for modulating metabolic inflammation and IR, we should address the role of specific immune cells and pro-inflammatory molecules in obesity and diabetes.

The cGAS–STING pathway was only thought to function in innate immune responses in early studies. However, as research progressed, it became evident that STING was expressed at high levels in CD4 + T cells in mice than in other immune cells [160, 161]. The STING pathway is thought to be critical in achieving anti-tumor responses in various tumor model systems [162–164]. The cGAS–STING pathway truly has a very important role not only in anti-tumor immunity but also in immune links to other diseases [25, 89]. For example, a recent study showed that the effects of *STING* mutations causing CD4 + and CD8 + T cell death in mice via ER stress and lung disease can be countered by altering T cell receptor specificity [165]. Thus, even though the various aspects of the cGAS–STING pathway in relation to immune cells and their mechanisms have not been fully elucidated, a large number of alternative propositions suggest that the cGAS–STING pathway affects disease processes via mediators other than only inflammatory factors [166]. There was no clear evidence of a link between diabetes and the cGAS–STING pathway mediated via immune cells until a recent study [167] confirmed a link between DsbA-L and T cells. Considering the link between DsbA-L and the cGAS–STING pathway, we may make some positive speculations, such as that DsbA-L might be a nexus between T cells and the cGAS–STING pathway. Zhou et al. [167], found that DsbA-L deficiency in HFD-fed mice suppressed mitochondrial function in T cells, and knocking down *DsbA-L* in specific T cells inhibited

IFN- γ production and accumulation of Treg cells, leading to reduced obesity in HFD-fed mice.

Combined with the performance of agonists of STING in NOD mice [24], the cGAS–STING pathway may inhibit the onset and progression of diabetes, despite the possible complex regulatory mechanisms. However, this evidence is inadequate because the mice in the NOD mice experiment suffered from type 1 diabetes, and there is no evidence to confirm that the cGAS–STING pathway is likewise able to inhibit the progression of diabetes in type 2 diabetes. In addition, NOD mice are just one of the type 1 diabetic mouse models and do not completely represent the full spectrum of type 1 diabetes. Therefore, although we lean toward the possibility that the cGAS–STING pathway may protect diabetic mice, the actual evidence is still not conclusive.

Potential link between the cGAS–STING pathway and diabetic complications

In addition to affecting energy metabolism, diabetes can also damage and/or inhibit the structure and function of several organs as the disease progresses, with diabetic cardiomyopathy and diabetic nephropathy being the most significant ones. Therefore, in this section, we will briefly discuss diabetes-induced complications and highlight the potential link between the cGAS–STING pathway and these diabetic complications.

Diabetic cardiomyopathy

Diabetic cardiomyopathy is an adverse consequence of the diabetes-related dysregulation of glucose and lipid metabolism affecting the heart, characterized by abnormalities in cardiac structure and function when other cardiovascular risk diagnoses have been excluded. It is likely to progress to heart failure [158]. Although the number of studies on diabetic cardiomyopathy has been increasing every year, the pathogenesis of this disease is still not well understood. In patients with diabetes, inflammatory cells in the heart secrete cytokines, chemokines, and exosomes that contribute to the development of myocardial hypertrophy, remodeling, and dysfunction. Inflammatory cells produce TNF, IL-6, IL-1 β , IFN- γ , and TGF- β cytokines, which induce or exacerbate myocardial injury, leading to further unfavorable remodeling and, consequently, diastolic and then systolic dysfunction [168–170]. NF- κ B directly induces cardiomyocyte hypertrophy and facilitates cardiac fibrosis [171]. Inflammatory pathways drive diabetic cardiomyopathy both directly and indirectly by increasing cardiomyocyte apoptosis and initiating signaling pathways, which leads to myocardial hypertrophy, fibrotic growth, and reduction or loss of cardiomyocyte contractility [158].

In addition to this, abnormal energy metabolism creates adverse conditions for the development of diabetic cardiomyopathy. The heart has a high energy demand, and abnormalities in energy metabolism will directly affect cardiac function. For example, glucose intake through GLUT4 requires the activation of the PI3K–Akt pathway to regulate translocation [172]. Thus, GLUT4 is required to maintain cardiac structure and function, and when Akt function is abnormal, it significantly affects the cardiac glucose oxidation rate, contributing to the development of diabetic cardiomyopathy [158, 172].

Previously, we described a potential link between Akt and the cGAS–STING pathway. Of note, one study showed that mice with *Akt2-AMPK* double-knockout developed cardiac dysfunction [154], which was consistent with previous reports that mice with single *Akt2* knockout develop typical T2DM [137, 173, 174]. In the double-knockout mice, cardiac dysfunction was adversely affected when *Akt2* and *AMPK* were knocked out, thus increasing the inflammatory responses after the activation of the cGAS–STING pathway [154]. Although there is only some indirect evidence, it is plausible that the cGAS–STING pathway exerts some influence on the development of diabetic cardiomyopathy, so we can make some degree of reasonable speculation that the cGAS–STING pathway promotes the development and progression of diabetic cardiomyopathy.

The treatment of endothelial cells with different concentrations of PA [88] or high levels of glucose [175] increased mitochondrial damage with mtDNA leakage into the cytoplasm and cGAS–STING pathway activation. Although endothelial cells are not cardiomyocytes, the contribution of cardiac endothelial dysfunction has been recognized and plays a major part in the pathogenesis of diabetic cardiomyopathy [177, 178]. A couple of earlier studies also showed that diabetes increased mtDNA damage and the glycation level of mtDNA repair enzyme [178, 179], which also indirectly indicates the potential activation of the cGAS–STING pathway owing to the presence of damaged mtDNA and mitochondria in the heart of diabetic mice. This assumption is also consistent with the above description of the essential role of the cGAS–STING signaling pathway in the pathogenesis of several other non-diabetic etiologies of cardiac remodeling and dysfunction [23, 90, 91, 93].

In summary, there is evidence that the cGAS–STING pathway may significantly contribute to the development of diabetes-related cardiac complications, and accurate modification of the cGAS–STING pathway of the heart may offer an essential contribution to the therapeutic and prognostic aspects of diabetic cardiomyopathy.

Association between the cGAS–STING pathway and the effects of diabetes in the liver

The liver plays important roles in metabolism, and exposure to various hepatic toxicants may damage the liver. Importantly, diabetes-mediated toxic effects also affect this organ [180]. It is well known that MetS or obesity commonly leads to NAFLD and hepatocyte lipotoxic injury, and the development of nonalcoholic steatohepatitis, which occur concurrently with the accumulation of insoluble protein aggregates composed of ubiquitinated proteins and the ubiquitin adaptor p62/sequestosome 1. Therefore, p62 inclusion formation in hepatocytes has been proposed as a pivotal marker to distinguish simple fatty liver from nonalcoholic steatohepatitis and possibly hepatocarcinogenesis with poor prognosis. Wang et al. reported that in response to lipotoxicity, TBK1 is triggered and phosphorylates p62, which is an important mediator of lipotoxicity-induced ubiquitinated protein aggregation and large protein inclusion formation in hepatocytes. These authors further confirmed that the inhibition of TBK1 both in cultured hepatocytes and mouse models of obesity and nonalcoholic steatohepatitis could prevent the formation of ubiquitin-p62 aggregates [181].

In summary, the above findings indicated that TBK1, a key node of the cGAS–STING pathway, is implicated in metabolic disease-induced liver disease, but there is still no convincing evidence that the cGAS–STING pathway as a whole is associated with diabetes-related liver dysfunction, which requires further study.

Diabetic nephropathy and the cGAS–STING pathway

Diabetic nephropathy is a common complication of diabetes and a major cause of chronic kidney disease [182]. Inflammation plays an equally critical role in the development and progression of diabetic nephropathy [183]. Hyperglycemia promotes the expression of inflammatory mediators (chemokines and cytokines) in injured glomeruli and tubular cells through different mechanisms of renal injury. These proinflammatory molecules also activate different signaling pathways such as the NF- κ B and JAK/STAT pathways [184–186]. They can also induce a cascade of responses, including vascular remodeling, endothelial dysfunction, extracellular matrix deposition, and mesenchymal proliferation [187, 188]. In addition to inflammatory responses, mitochondrial dysfunction can cause chronic kidney disease [189]. In the kidney, high plasma glucose causes immediate tubular cell damage, leading to extensive metabolic and cellular dysfunction. In particular, adenine nucleotides, specifically ATP, are essential metabolic modulators of tubular glomerular

responses in the kidney, which influence both the renal blood stream and the quantity of oxygen delivered [190]. In contrast, in diabetic nephropathy, alterations in extramural ATP via type 2 purinergic receptors already occur in the early stages of the disease, and such changes cause interstitial fibrillation [191, 192]. Abnormal mitochondrial metabolism has a major role in diabetic nephropathy, and the cGAS–STING pathway has been confirmed to be activated by abnormal mitochondrial function. Therefore, we can speculate that the cGAS–STING signaling pathway may participate in the pathogenesis of diabetic nephropathy, and the exploration of this mechanism may help elucidate the pathophysiology of diabetic nephropathy.

Conclusion

It is widely documented that inflammation and impaired mitochondrial function promote disease processes and that the cGAS–STING pathway can potentially partake in inflammation and impaired mitochondrial function. Therefore, investigation on the cGAS–STING pathway could be extended to more diseases involving inflammation. At the same time, owing to the contribution of the cGAS–STING pathway in different organs, inhibition of the cGAS–STING pathway-induced inflammatory pathway provides a potential avenue for therapeutic drug development for diabetes and subsequent diabetes-related complications. Although researchers have investigated the cGAS–STING approach, there are still many critical aspects to be addressed during the development of practical applications. First, although the cGAS–STING pathway has been studied extensively, the main focus of the studies has been resistance to various microbial pathogens, whereas the actions of this pathway have not yet been investigated in non-immune cells. Second, the cGAS–STING pathway may respond differently to different environments, given that different environments will have different substantial effects on the organism; for example, whether the cGAS–STING pathway is activated in mice exposed to high manganese; however, there is no literature investigating the effects of environmental changes on the cGAS–STING pathway. Lastly, is it possible to treat diabetes by targeting the cGAS–STING pathway? This is not only with regards to the possibility of targeted therapy but also to the implementation of targeted therapy. We believe that as systemic and detailed approaches have been recommended for monitoring cGAS–STING signaling in cultured cells and tissues of mice [193, 194], studies on the cGAS–STING pathway in diabetes and the ensuing complications are promising and feasible, and we intend to investigate the abovementioned topics to some extent in future studies.

Abbreviations

AECOPD: Acute exacerbation of COPD; ALD: Alcoholic liver disease; AMPK: AMP-activated protein kinase; BAX: BCL-2 related X protein; COPD: Chronic obstructive pulmonary disease; cGAMP: Cyclic guanosine monophosphate-adenosine monophosphate; CMV: Cytomegalovirus; cGAS: Cytosolic Gmp-amp synthase; DNase II: Deoxyribonuclease II; DM: Diabetes mellitus; DCM: Dilated cardiomyopathy; DsbA-L: Disulfide bond A oxidoreductase-like protein; DSBs: DNA double-strand breaks; HSV-1: DNA herpes simplex virus type 1; dsDNA: Double-stranded DNA; ER: Endoplasmic reticulum; EBV: Epstein Barr virus; ERGIC: ER-Golgi intermediate; GLUT-4: Glucose transporter-4; HBV: Hepatitis B virus; HFD: High fat diet; HDM: House dust mite; HIV: Human immunodeficiency virus; HCM: Hypertrophic cardiomyopathy; iNOS: Inducible nitric oxide synthase; IR: Insulin resistance; IFN: Interferon; IRF3: Interferon regulatory factor 3; ISG: Interferon-stimulated gene; LPS: Lipopolysaccharide; LN: Lupus nephritis; MetS: Metabolic syndrome; mtDNA: Mitochondrial DNA; MLKL: Mixed lineage kinase domain-like pseudokinase; mTORC1: MTOR complex 1; mTORC2: MTOR complex 2; NLRP3: NLR family pyrin domain containing 3; NAFLD: Non-alcoholic fatty liver disease; NF-κB: Nuclear factor kappa-light-chain-enhancer of activated B cells; PA: Palmitic acid; PPARs: Peroxisome proliferator-activated receptor; PKB/Akt: Protein kinase B; RIP3: Receptor-interacting serine/threonine kinase 3; rAdVs: Recombinant adenovirus vector; RA: Rheumatoid arthritis; SeV: Sendai virus; SASP: Senescence-associated secretory phenotype; STAT: Signal transducer and activator of transcription; STING: Stimulator of interferon gene; SAVI: STING-associated vasculopathy with onset in infancy; SLE: Systemic lupus erythematosus; TBK1: TANK-binding kinase 1; mTOR: The mammalian target of rapamycin; PI3-K: The phosphatidylyl 3 kinase; TNF-α: Tumor necrosis factor alpha; T1DM: Type 1 diabetes mellitus; T2DM: Type 2 diabetes mellitus; VAPS: Vascular and pulmonary syndrome; VSV: Vesicular stomatitis; WAT: White adipose tissue.

Acknowledgements

This work was partially supported by the China Jilin Province Science and Technology Department Project (20190701067GH). The authors are grateful to the U.S.-China Pediatric Research Exchange Training Program. The personnel expenses for T Bao when he was trained in the University of Louisville (3,2019–5,2021) was provided by the First Hospital of Jilin University, Changchun, China. We also are indebted to Editage (www.editage.cn) for the English writing editing.

Authors' contributions

TB performed literature sorting and wrote the article; LJ collected literature and wrote the article; JL and LC conceived and designed the study and drafted and reviewed the manuscript. All authors read and approved the final manuscript.

Funding

This work was supported in part by the China Jilin Province Science and Technology Department Project (20190701067GH).

Availability of data and materials

Not applicable.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Its publication has been approved by all co-authors.

Competing interests

The authors declare that they have no competing interests.

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Received: 30 April 2021 Accepted: 2 December 2021

Published online: 14 December 2021

References

1. Third Report of the National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (Adult Treatment Panel III) final report. *Circulation*. 2002;106(25):3143–421. <https://doi.org/10.1161/circ.106.25.3143>
2. Pararasa C, Bailey CJ, Griffiths HR. Ageing, adipose tissue, fatty acids and inflammation. *Biogerontology*. 2015;16(2):235–48. <https://doi.org/10.1007/s10522-014-9536-x>.
3. Hotamisligil GS. Inflammation and metabolic disorders. *Nature*. 2006;444(7121):860–7. <https://doi.org/10.1038/nature05485>.
4. Eguchi K, Nagai R. Islet inflammation in type 2 diabetes and physiology. *J Clin Invest*. 2017;127(1):14–23. <https://doi.org/10.1172/JCI88877>.
5. DiMeglio LA, Evans-Molina C, Oram RA. Type 1 diabetes. *Lancet*. 2018;391(10138):2449–62. [https://doi.org/10.1016/S0140-6736\(18\)31320-5](https://doi.org/10.1016/S0140-6736(18)31320-5).
6. Donath MY, et al. Islet inflammation impairs the pancreatic beta-cell in type 2 diabetes. *Physiology (Bethesda)*. 2009;24:325–31. <https://doi.org/10.1152/physiol.00032.2009>.
7. Chung IM, et al. Insights on the current status and advancement of diabetes mellitus type 2 and to avert complications: an overview. *Biotechnol Appl Biochem*. 2020;67(6):920–8. <https://doi.org/10.1002/bab.1853>.
8. Chen Q, Sun L, Chen ZJ. Regulation and function of the cGAS–STING pathway of cytosolic DNA sensing. *Nat Immunol*. 2016;17(10):1142–9. <https://doi.org/10.1038/ni.3558>.
9. Tan X, et al. Detection of microbial infections through innate immune sensing of nucleic acids. *Annu Rev Microbiol*. 2018;72:447–78. <https://doi.org/10.1146/annurev-micro-102215-095605>.
10. Kwon J, Bakhom SF. The cytosolic DNA-sensing cGAS–STING pathway in cancer. *Cancer Discov*. 2020;10(1):26–39. <https://doi.org/10.1158/2159-8290.CD-19-0761>.
11. Jiang M, et al. cGAS–STING, an important pathway in cancer immunotherapy. *J Hematol Oncol*. 2020;13(1):81. <https://doi.org/10.1186/s13045-020-00916-z>.
12. Kumar V. A STING to inflammation and autoimmunity. *J Leukoc Biol*. 2019;106(1):171–85. <https://doi.org/10.1002/JLB.4MIR1018-397RR>.
13. Bai J, Liu F. The cGAS–cGAMP–STING pathway: a molecular link between immunity and metabolism. *Diabetes*. 2019;68(6):1099–108. <https://doi.org/10.2337/dbi18-0052>.
14. Bai J, et al. DsbA-L prevents obesity-induced inflammation and insulin resistance by suppressing the mtDNA release-activated cGAS–cGAMP–STING pathway. *Proc Natl Acad Sci U S A*. 2017;114(46):12196–201. <https://doi.org/10.1073/pnas.1708744114>.
15. Kranzusch PJ, et al. Ancient origin of cGAS–STING reveals mechanism of universal 2',3' cGAMP signaling. *Mol Cell*. 2015;59(6):891–903. <https://doi.org/10.1016/j.molcel.2015.07.022>.
16. Ishikawa H, Barber GN. STING is an endoplasmic reticulum adaptor that facilitates innate immune signalling. *Nature*. 2008;455(7213):674–8. <https://doi.org/10.1038/nature07317>.
17. Zhong B, et al. The adaptor protein MITA links virus-sensing receptors to IRF3 transcription factor activation. *Immunity*. 2008;29(4):538–50. <https://doi.org/10.1016/j.immuni.2008.09.003>.
18. Jin L, et al. MPYS, a novel membrane tetraspanner, is associated with major histocompatibility complex class II and mediates transduction of apoptotic signals. *Mol Cell Biol*. 2008;28(16):5014–26. <https://doi.org/10.1128/MCB.00640-08>.
19. Sun L, et al. Cyclic GMP-AMP synthase is a cytosolic DNA sensor that activates the type I interferon pathway. *Science (American Association for the Advancement of Science)*. 2012;339(6121):786–91. <https://doi.org/10.1126/science.1232458>.
20. Wu J, Sun L, Chen X, Du F, Shi H, Chen C, Chen ZJ. Cyclic GMP-AMP is an endogenous second messenger in innate immune signaling by cytosolic DNA. *Science*. 2013;339:826–30. <https://doi.org/10.1126/science.1229963>.
21. Qiao JT, et al. Activation of the STING–IRF3 pathway promotes hepatocyte inflammation, apoptosis and induces metabolic disorders in

- nonalcoholic fatty liver disease. *Metabolism*. 2018;81:13–24. <https://doi.org/10.1016/j.metabol.2017.09.010>.
22. Schuliga M, et al. Self DNA perpetuates IPF lung fibroblast senescence in a cGAS-dependent manner. *Clin Sci (Lond)*. 2020;134(7):889–905.
 23. Hu D, et al. Cytosolic DNA sensor cGAS plays an essential pathogenetic role in pressure overload-induced heart failure. *Am J Physiol Heart Circ Physiol*. 2020;318(6):H1525–37. <https://doi.org/10.1152/ajpheart.00097.2020>.
 24. Lemos H, et al. Stimulator of interferon genes agonists attenuate type I diabetes progression in NOD mice. *Immunology*. 2019;158(4):353–61. <https://doi.org/10.1111/imm.13122>.
 25. Cao DJ, et al. Cytosolic DNA sensing promotes macrophage transformation and governs myocardial ischemic injury. *Circulation*. 2018;137(24):2613–34. <https://doi.org/10.1161/CIRCULATIONAHA.117.031046>.
 26. Li X, et al. Cyclic GMP-AMP synthase is activated by double-stranded DNA-induced oligomerization. *Immunity*. 2013;39(6):1019–31. <https://doi.org/10.1016/j.immuni.2013.10.019>.
 27. Zhang X, et al. The cytosolic DNA sensor cGAS forms an oligomeric complex with DNA and undergoes switch-like conformational changes in the activation loop. *Cell Rep*. 2014;6(3):421–30. <https://doi.org/10.1016/j.celrep.2014.01.003>.
 28. Andreeva L, et al. cGAS senses long and HMGB/TFAM-bound U-turn DNA by forming protein–DNA ladders. *Nature*. 2017;549(7672):394–8. <https://doi.org/10.1038/nature23890>.
 29. Hooy RM, Sohn J. The allosteric activation of cGAS underpins its dynamic signaling landscape. *Elife*. 2018. <https://doi.org/10.7554/eLife.39984>.
 30. West AP, et al. Mitochondrial DNA stress primes the antiviral innate immune response. *Nature*. 2015;520(7548):553–7. <https://doi.org/10.1038/nature14156>.
 31. Mackenzie KJ, et al. cGAS surveillance of micronuclei links genome instability to innate immunity. *Nature*. 2017;548(7668):461–5. <https://doi.org/10.1038/nature23449>.
 32. Harding SM, et al. Mitotic progression following DNA damage enables pattern recognition within micronuclei. *Nature*. 2017;548(7668):466–70. <https://doi.org/10.1038/nature23470>.
 33. Dou Z, et al. Cytoplasmic chromatin triggers inflammation in senescence and cancer. *Nature*. 2017;550(7676):402–6. <https://doi.org/10.1038/nature24050>.
 34. Gratia M, et al. Bloom syndrome protein restrains innate immune sensing of micronuclei by cGAS. *J Exp Med*. 2019;216(5):1199–213. <https://doi.org/10.1084/jem.20181329>.
 35. Ablasser A, et al. cGAS produces a 2'-5'-linked cyclic dinucleotide second messenger that activates STING. *Nature*. 2013;498(7454):380–4. <https://doi.org/10.1038/nature12306>.
 36. Kranzusch PJ, et al. Structure-guided reprogramming of human cGAS dinucleotide linkage specificity. *Cell*. 2014;158(5):1011–21. <https://doi.org/10.1016/j.cell.2014.07.028>.
 37. Liu H, et al. Nuclear cGAS suppresses DNA repair and promotes tumorigenesis. *Nature*. 2018;563(7729):131–6. <https://doi.org/10.1038/s41586-018-0629-6>.
 38. Jiang H, et al. Chromatin-bound cGAS is an inhibitor of DNA repair and hence accelerates genome destabilization and cell death. *EMBO J*. 2019;38(21):e102718. <https://doi.org/10.15252/embj.2019102718>.
 39. Wang Y, et al. Inflammasome activation triggers caspase-1-mediated cleavage of cGAS to regulate responses to DNA virus infection. *Immunity*. 2017;46(3):393–404. <https://doi.org/10.1016/j.immuni.2017.02.011>.
 40. Ning X, et al. Apoptotic caspases suppress type I interferon production via the cleavage of cGAS, MAVS, and IRF3. *Mol Cell*. 2019;74(1):19–31.e7. <https://doi.org/10.1016/j.molcel.2019.02.013>.
 41. Seo GJ, et al. Akt kinase-mediated checkpoint of cGAS DNA sensing pathway. *Cell Rep*. 2015;13(2):440–9. <https://doi.org/10.1016/j.celrep.2015.09.007>.
 42. Cui Y, et al. SENP7 potentiates cGAS activation by relieving SUMO-mediated inhibition of cytosolic DNA sensing. *PLoS Pathog*. 2017;13(1):e1006156. <https://doi.org/10.1371/journal.ppat.1006156>.
 43. Hu M, et al. Sumoylation promotes the stability of the DNA sensor cGAS and the adaptor STING to regulate the kinetics of response to DNA virus. *Immunity*. 2016;45(3):555–69. <https://doi.org/10.1016/j.immuni.2016.08.014>.
 44. Xia P, et al. Glutamylation of the DNA sensor cGAS regulates its binding and synthase activity in antiviral immunity. *Nat Immunol*. 2016;17(4):369–78. <https://doi.org/10.1038/ni.3356>.
 45. Dai J, et al. Acetylation blocks cGAS activity and inhibits self-DNA-induced autoimmunity. *Cell*. 2019;176(6):1447–1460.e14. <https://doi.org/10.1016/j.cell.2019.01.016>.
 46. Chen M, et al. TRIM14 inhibits cGAS degradation mediated by selective autophagy receptor p62 to promote innate immune responses. *Mol Cell*. 2016;64(1):105–19. <https://doi.org/10.1016/j.molcel.2016.08.025>.
 47. Wang Q, et al. The E3 ubiquitin ligase RNF185 facilitates the cGAS-mediated innate immune response. *PLoS Pathog*. 2017;13(3):e1006264. <https://doi.org/10.1371/journal.ppat.1006264>.
 48. Guo Y, et al. Cutting edge: USP27X deubiquitinates and stabilizes the DNA sensor cGAS to regulate cytosolic DNA-mediated signaling. *J Immunol*. 2019;203(8):2049–54. <https://doi.org/10.4049/jimmunol.1900514>.
 49. Shang G, et al. Cryo-EM structures of STING reveal its mechanism of activation by cyclic GMP–AMP. *Nature*. 2019;567(7748):389–93. <https://doi.org/10.1038/s41586-019-0998-5>.
 50. Zhang C, et al. Structural basis of STING binding with and phosphorylation by TBK1. *Nature*. 2019;567(7748):394–8. <https://doi.org/10.1038/s41586-019-1000-2>.
 51. Hiscott J, et al. Triggering the interferon response: the role of IRF-3 transcription factor. *J Interferon Cytokine Res*. 1999;19(1):1–13. <https://doi.org/10.1089/107999099314360>.
 52. Dobbs N, et al. STING activation by translocation from the ER is associated with infection and autoinflammatory disease. *Cell Host Microbe*. 2015;18(2):157–68. <https://doi.org/10.1016/j.chom.2015.07.001>.
 53. de Oliveira MC, et al. Modular architecture of the STING C-terminal tail allows interferon and NF-kappaB signaling adaptation. *Cell Rep*. 2019;27(4):1165–1175.e5. <https://doi.org/10.1016/j.celrep.2019.03.098>.
 54. Liu S, et al. Phosphorylation of innate immune adaptor proteins MAVS, STING, and TRIF induces IRF3 activation. *Science*. 2015;347(6227):aaa2630. <https://doi.org/10.1126/science.aaa2630>.
 55. Tanaka Y, Chen ZJ. STING specifies IRF3 phosphorylation by TBK1 in the cytosolic DNA signaling pathway. *Sci Signal*. 2012;5(214):ra20. <https://doi.org/10.1126/scisignal.2002521>.
 56. Gonzalez-Navajas JM, et al. Immunomodulatory functions of type I interferons. *Nat Rev Immunol*. 2012;12(2):125–35. <https://doi.org/10.1038/nri3133>.
 57. Andrienas KK, et al. DNA-binding landscape of IRF3, IRF5 and IRF7 dimers: implications for dimer-specific gene regulation. *Nucleic Acids Res*. 2018;46(5):2509–20. <https://doi.org/10.1093/nar/gky002>.
 58. Schoggins JW, et al. A diverse range of gene products are effectors of the type I interferon antiviral response. *Nature*. 2011;472(7344):481–5. <https://doi.org/10.1038/nature09907>.
 59. Watson RO, Manzanillo PS, Cox JS. Extracellular *M. tuberculosis* DNA targets bacteria for autophagy by activating the host DNA-sensing pathway. *Cell*. 2012;150(4):803–15. <https://doi.org/10.1016/j.cell.2012.06.040>.
 60. Liu F, et al. Beclin1 haploinsufficiency accentuates second-hand smoke exposure-induced myocardial remodeling and contractile dysfunction through a STING-mediated mechanism. *J Mol Cell Cardiol*. 2020;148:78–88. <https://doi.org/10.1016/j.yjmcc.2020.08.016>.
 61. Abdullah A, et al. STING-mediated autophagy is protective against H₂O₂-induced cell death. *Int J Mol Sci*. 2020. <https://doi.org/10.3390/ijms21197059>.
 62. Glück S, et al. Innate immune sensing of cytosolic chromatin fragments through cGAS promotes senescence. *Nat Cell Biol*. 2017;19(9):1061–70. <https://doi.org/10.1038/ncb3586>.
 63. Coppé J, et al. The senescence-associated secretory phenotype: the dark side of tumor suppression. *Annu Rev Pathol*. 2010;5(1):99–118. <https://doi.org/10.1146/annurev-pathol-121808-102144>.
 64. Xue W, et al. Senescence and tumour clearance is triggered by p53 restoration in murine liver carcinomas. *Nature*. 2007;445(7128):656–60. <https://doi.org/10.1038/nature05529>.
 65. Gulen MF, et al. Signalling strength determines proapoptotic functions of STING. *Nat Commun*. 2017. <https://doi.org/10.1038/s41467-017-00573-w>.
 66. Paludan SR, Reinert LS, Hornung V. DNA-stimulated cell death: implications for host defence, inflammatory diseases and cancer.

- Nat Rev Immunol. 2019;19(3):141–53. <https://doi.org/10.1038/s41577-018-0117-0>.
67. Chen D, et al. PUMA amplifies necroptosis signaling by activating cytosolic DNA sensors. *Proc Natl Acad Sci*. 2018;115(15):3930–5. <https://doi.org/10.1073/pnas.1717190115>.
 68. Sze A, et al. Host restriction factor SAMHD1 limits human T cell leukemia virus type 1 infection of monocytes via STING-mediated apoptosis. *Cell Host Microbe*. 2013;14(4):422–34. <https://doi.org/10.1016/j.chom.2013.09.009>.
 69. Gaidt MM, et al. The DNA inflammasome in human myeloid cells is initiated by a STING-cell death program upstream of NLRP3. *Cell*. 2017;171(5):1110–1124.e18. <https://doi.org/10.1016/j.cell.2017.09.039>.
 70. Motwani M, Pawaria S, Bernier J. Hierarchy of clinical manifestations in SAVI N153S and V154M mouse models. *Proc Natl Acad Sci*. 2019;116(16):7941–50. <https://doi.org/10.1073/pnas.1818281116>.
 71. Kalamvoki M, Roizman B. HSV-1 degrades, stabilizes, requires, or is stung by STING depending on ICP0, the US3 protein kinase, and cell derivation. *Proc Natl Acad Sci*. 2014;111(5):E611–7. <https://doi.org/10.1073/pnas.1323414111>.
 72. Guo F, et al. Activation of stimulator of interferon genes in hepatocytes suppresses the replication of hepatitis B virus. *Antimicrob Agents Chemother*. 2017. <https://doi.org/10.1128/AAC.00771-17>.
 73. Chen Q, et al. Carcinoma-astrocyte gap junctions promote brain metastasis by cGAMP transfer. *Nature*. 2016;533(7604):493–8. <https://doi.org/10.1038/nature18268>.
 74. Liu Y, et al. Activated STING in a vascular and pulmonary syndrome. *N Engl J Med*. 2014;371(6):507–18. <https://doi.org/10.1056/NEJMoa1312625>.
 75. Picard C, et al. Severe pulmonary fibrosis as the first manifestation of interferonopathy (TMEM173 mutation). *Chest*. 2016;150(3):e65–71. <https://doi.org/10.1016/j.chest.2016.02.682>.
 76. Seo J, et al. Tofacitinib relieves symptoms of stimulator of interferon genes (STING)-associated vasculopathy with onset in infancy caused by 2 de novo variants in TMEM173. *J Allergy Clin Immunol*. 2017;139(4):1396–1399.e12. <https://doi.org/10.1016/j.jaci.2016.10.030>.
 77. Melki I, et al. Disease-associated mutations identify a novel region in human STING necessary for the control of type I interferon signaling. *J Allergy Clin Immunol*. 2017;140(2):543–552.e5. <https://doi.org/10.1016/j.jaci.2016.10.031>.
 78. Gall A, et al. Autoimmunity initiates in nonhematopoietic cells and progresses via lymphocytes in an interferon-dependent autoimmune disease. *Immunity*. 2012;36(1):120–31. <https://doi.org/10.1016/j.immuni.2011.11.018>.
 79. Crow YJ, et al. Mutations in the gene encoding the 3'-5' DNA exonuclease TREX1 cause Aicardi-Goutières syndrome at the AGS1 locus. *Nat Genet*. 2006;38(8):917–20.
 80. Konno H, et al. Pro-inflammation associated with a gain-of-function mutation (R284S) in the innate immune sensor STING. *Cell Rep*. 2018;23(4):1112–23. <https://doi.org/10.1038/ng1845>.
 81. An J, et al. Expression of cyclic GMP-AMP synthase in patients with systemic lupus erythematosus. *Arthritis Rheumatol*. 2017;69(4):800–7. <https://doi.org/10.1002/art.40002>.
 82. Jeremiah N, et al. Inherited STING-activating mutation underlies a familial inflammatory syndrome with lupus-like manifestations. *J Clin Invest*. 2014;124(12):5516–20. <https://doi.org/10.1172/JCI79100>.
 83. Gehrke N, Mertens C, Zillinger T. Oxidative damage of DNA confers resistance to cytosolic nuclease TREX1 degradation and potentiates STING-dependent immune sensing. *Immunity*. 2013;39(3):482–95. <https://doi.org/10.1016/j.immuni.2013.08.004>.
 84. Yang Y, Lindahl T, Barnes DE. Trex1 exonuclease degrades ssDNA to prevent chronic checkpoint activation and autoimmune disease. *Cell*. 2007;131(5):873–86. <https://doi.org/10.1016/j.cell.2007.10.017>.
 85. Morita M, et al. Gene-targeted mice lacking the Trex1 (DNase III) 3'→5' DNA exonuclease develop inflammatory myocarditis. *Mol Cell Biol*. 2004;24(15):6719–27. <https://doi.org/10.1128/MCB.24.15.6719>.
 86. Gao D, et al. Activation of cyclic GMP-AMP synthase by self-DNA causes autoimmune diseases. *Proc Natl Acad Sci*. 2015;112(42):E5699–705. <https://doi.org/10.1073/pnas.1516465112>.
 87. Ding L, et al. The regional function of cGAS/STING signal in multiple organs: One of culprit behind systemic lupus erythematosus? *Med Hypotheses*. 2015;85(6):846–9. <https://doi.org/10.1016/j.mehy.2015.09.026>.
 88. Mao Y, et al. STING-IRF3 triggers endothelial inflammation in response to free fatty acid-induced mitochondrial damage in diet-induced obesity. *Arterioscler Thromb Vasc Biol*. 2017;37(5):920–9. <https://doi.org/10.1161/ATVBAHA.117.309017>.
 89. King KR, et al. IRF3 and type I interferons fuel a fatal response to myocardial infarction. *Nat Med*. 2017;23(12):1481–7. <https://doi.org/10.1038/nm.4428>.
 90. Li N, et al. STING-IRF3 contributes to lipopolysaccharide-induced cardiac dysfunction, inflammation, apoptosis and pyroptosis by activating NLRP3. *Redox Biol*. 2019;24: 101215. <https://doi.org/10.1016/j.redox.2019.101215>.
 91. Xu Q, et al. Small molecule inhibition of cyclic GMP-AMP synthase ameliorates sepsis-induced cardiac dysfunction in mice. *Life Sci*. 2020;260: 118315. <https://doi.org/10.1016/j.lfs.2020.118315>.
 92. Wang S, et al. ALDH2 contributes to melatonin-induced protection against APP/PS1 mutation-prompted cardiac anomalies through cGAS-STING-TBK1-mediated regulation of mitophagy. *Signal Transduct Target Ther*. 2020;5(1):119. <https://doi.org/10.1038/s41392-020-0171-5>.
 93. Zhang Y, Chen W, Wang Y. STING is an essential regulator of heart inflammation and fibrosis in mice with pathological cardiac hypertrophy via endoplasmic reticulum (ER) stress. *Biomed Pharmacother*. 2020;125: 110022. <https://doi.org/10.1016/j.biopha.2020.110022>.
 94. Wang J, et al. Cardiac overexpression of 8-oxoguanine DNA glycosylase 1 protects mitochondrial DNA and reduces cardiac fibrosis following transaortic constriction. *Am J Physiol Heart Circ Physiol*. 2011;301(5):H2073–80. <https://doi.org/10.1152/ajpheart.00157.2011>.
 95. Calcagno DM, et al. The myeloid type I interferon response to myocardial infarction begins in bone marrow and is regulated by Nrf2-activated macrophages. *Sci Immunol*. 2020. <https://doi.org/10.1126/sciimmunol.aaz1974>.
 96. Hilzenderger C, et al. Reduced sputum expression of interferon-stimulated genes in severe COPD. *Int J Chron Obstruct Pulmon Dis*. 2016;11:1485–94. <https://doi.org/10.2147/COPD.S105948>.
 97. Nascimento M, et al. Self-DNA release and STING-dependent sensing drives inflammation to cigarette smoke in mice. *Sci Rep*. 2019;9(1):14848. <https://doi.org/10.1038/s41598-019-51427-y>.
 98. Qin H, et al. Diminished stimulator of interferon genes production with cigarette smoke-exposure contributes to weakened anti-adenovirus vectors response and destruction of lung in chronic obstructive pulmonary disease model. *Exp Cell Res*. 2019;384(1): 111545. <https://doi.org/10.1016/j.yexcr.2019.111545>.
 99. Benmerzoug S, et al. STING-dependent sensing of self-DNA drives silica-induced lung inflammation. *Nat Commun*. 2018;9(1):5226. <https://doi.org/10.1038/s41467-018-07425-1>.
 100. Qiu H, et al. Stimulator of interferon genes deficiency in acute exacerbation of idiopathic pulmonary fibrosis. *Front Immunol*. 2017;8:1756. <https://doi.org/10.3389/fimmu.2017.01756>.
 101. Savigny F, et al. Protective role of the nucleic acid sensor STING in pulmonary fibrosis. *Front Immunol*. 2020;11: 588799. <https://doi.org/10.3389/fimmu.2020.588799>.
 102. Han Y, et al. Airway epithelial cGAS is critical for induction of experimental allergic airway inflammation. *J Immunol*. 2020;204(6):1437–47. <https://doi.org/10.4049/jimmunol.1900869>.
 103. Bhakta NR, et al. IFN-stimulated gene expression, type 2 inflammation, and endoplasmic reticulum stress in asthma. *Am J Respir Crit Care Med*. 2018;197(3):313–24. <https://doi.org/10.1164/rccm.201706-10700C>.
 104. Yu Y, Liu Y, An W, Song J, Zhang Y, Zhao X. STING-mediated inflammation in Kupffer cells contributes to progression of nonalcoholic steatohepatitis. *J Clin Invest*. 2019;129:546–55. <https://doi.org/10.1172/JCI121842>.
 105. Luo X, Li H, Ma L, Zhou J, Guo X, Woo S-L, et al. Expression of STING is increased in liver tissues from patients with NAFLD and promotes macrophage-mediated hepatic inflammation and fibrosis in mice. *Gastroenterology*. 2018;155:1971–84. <https://doi.org/10.1053/j.gastro.2018.09.010>.
 106. Wang X, et al. STING expression in monocyte-derived macrophages is associated with the progression of liver inflammation and fibrosis in patients with nonalcoholic fatty liver disease. *Lab Invest*. 2020;100(4):542–52. <https://doi.org/10.1038/s41374-019-0342-6>.

107. Iracheta-Vellve A, et al. Endoplasmic reticulum stress-induced hepatocellular death pathways mediate liver injury and fibrosis via stimulator of interferon genes. *J Biol Chem*. 2016;291(52):26794–805. <https://doi.org/10.1074/jbc.M116.736991>.
108. Petrasek J, et al. STING-IRF3 pathway links endoplasmic reticulum stress with hepatocyte apoptosis in early alcoholic liver disease. *Proc Natl Acad Sci*. 2013;110(41):16544–9. <https://doi.org/10.1073/pnas.1308331110>.
109. Luther J, et al. Hepatic gap junctions amplify alcohol liver injury by propagating cGAS-mediated IRF3 activation. *Proc Natl Acad Sci U S A*. 2020;117(21):11667–73. <https://doi.org/10.1073/pnas.1911870117/-/DCSupplemental>.
110. Lei Z, et al. cGAS-mediated autophagy protects the liver from ischemia-reperfusion injury independently of STING. *Am J Physiol Gastrointest Liver Physiol*. 2018;314(6):G655–67. <https://doi.org/10.1152/ajpgi.00326.2017>.
111. Zhong W, et al. Aging aggravated liver ischemia and reperfusion injury by promoting STING-mediated NLRP3 activation in macrophages. *Aging Cell*. 2020;19(8):e13186. <https://doi.org/10.1111/acer.13186>.
112. Xue L, et al. Tumor necrosis factor-like weak inducer of apoptosis activates type I interferon signals in lupus nephritis. *Biomed Res Int*. 2017;2017:4927376. <https://doi.org/10.1155/2017/4927376>.
113. Maekawa H, et al. Mitochondrial damage causes inflammation via cGAS–STING signaling in acute kidney injury. *Cell Rep*. 2019;29(5):1261–1273.e6. <https://doi.org/10.1016/j.celrep.2019.09.050>.
114. Chung KW, et al. Mitochondrial damage and activation of the STING pathway lead to renal inflammation and fibrosis. *Cell Metab*. 2019;30(4):784–799.e5. <https://doi.org/10.1016/j.cmet.2019.08.003>.
115. Davis SE, Khatua AK, Popik W. Nucleosomal dsDNA stimulates APOL1 expression in human cultured podocytes by activating the cGAS/IF116-STING signaling pathway. *Sci Rep*. 2019;9(1):15485. <https://doi.org/10.1038/s41598-019-51998-w>.
116. Barrett JP, Henry RJ, et al. Interferon- β plays a detrimental role in experimental traumatic brain injury by enhancing neuroinflammation that drives chronic neurodegeneration. *J Neurosci*. 2020;40(11):2357–70. <https://doi.org/10.1523/JNEUROSCI.2516-19.2020>.
117. Li Q, et al. Inhibition of double-strand DNA-sensing cGAS ameliorates brain injury after ischemic stroke. *EMBO Mol Med*. 2020;12(4):e11002. <https://doi.org/10.15252/emmm.201911002>.
118. Sharma M, et al. Cyclic GMP-AMP synthase promotes the inflammatory and autophagy responses in Huntington disease. *Proc Natl Acad Sci U S A*. 2020;117(27):15989–99. <https://doi.org/10.1073/pnas.2002144117/-/DCSupplemental>.
119. Kerur N, et al. cGAS drives noncanonical-inflammasome activation in age-related macular degeneration. *Nat Med*. 2018;24(1):50–61. <https://doi.org/10.1038/nm.4450>.
120. Luo W, et al. Critical role of cytosolic DNA and its sensing adaptor STING in aortic degeneration, dissection, and rupture. *Circulation*. 2020;141(1):42–66. <https://doi.org/10.1161/CIRCULATIONAHA.119.041460>.
121. Yuan L, et al. Palmitic acid dysregulates the Hippo–YAP pathway and inhibits angiogenesis by inducing mitochondrial damage and activating the cytosolic DNA sensor cGAS–STING–IRF3 signaling mechanism. *J Biol Chem*. 2017;292(36):15002–15. <https://doi.org/10.1074/jbc.M117.804005>.
122. Zhao Q, et al. STING signaling promotes inflammation in experimental acute pancreatitis. *Gastroenterology*. 2018;154(6):1822–1835.e2. <https://doi.org/10.1053/j.gastro.2018.01.065>.
123. Ma C, et al. Gasdermin D in macrophages restrains colitis by controlling cGAS-mediated inflammation. *Sci Adv*. 2020;6(21):eaa26717. <https://doi.org/10.1126/sciadv.aaz6717>.
124. Association AD. 1. Improving care and promoting health in populations: standards of medical care in diabetes-2021. *Diabetes Care*. 2021;44(Suppl 1):7–14. <https://doi.org/10.2337/dc21-S001>.
125. Phillips JM, et al. Type 1 diabetes development requires both CD4+ and CD8+ T cells and can be reversed by non-depleting antibodies targeting both T cell populations. *Rev Diabet Stud*. 2009;6(2):97–103. <https://doi.org/10.1900/RDS.2009.6.97>.
126. Richardson SJ, Morgan NG, Foulis AK. Pancreatic pathology in type 1 diabetes mellitus. *Endocr Pathol*. 2014;25(1):80–92. <https://doi.org/10.1007/s12022-014-9297-8>.
127. Tsalamandris S, et al. The role of inflammation in diabetes: current concepts and future perspectives. *Eur Cardiol*. 2019;14(1):50–9. <https://doi.org/10.15420/ecr.2018.33.1>.
128. DeFronzo RA. Pathogenesis of type 2 diabetes mellitus. *Med Clin North Am*. 2004;88(4):787–835, ix. <https://doi.org/10.1016/j.mcna.2004.04.013>.
129. Lumeng CN, Saltiel AR. Inflammatory links between obesity and metabolic disease. *J Clin Invest*. 2011;121(6):2111–7. <https://doi.org/10.1172/JCI57132>.
130. Shoelson SE, Herrero L, Naaz A. Obesity, inflammation, and insulin resistance. *Gastroenterology*. 2007;132(6):2169–80. <https://doi.org/10.1053/j.gastro.2007.03.059>.
131. Kanda H, et al. MCP-1 contributes to macrophage infiltration into adipose tissue, insulin resistance, and hepatic steatosis in obesity. *J Clin Invest*. 2006;116(6):1494–505. <https://doi.org/10.1172/JCI26498>.
132. Antoniadou C, et al. Adiponectin: from obesity to cardiovascular disease. *Obes Rev*. 2009;10(3):269–79. <https://doi.org/10.1111/j.1467-789X.2009.00571.x>.
133. Antonopoulos AS, et al. Adiponectin as a link between type 2 diabetes and vascular NADPH oxidase activity in the human arterial wall: the regulatory role of perivascular adipose tissue. *Diabetes*. 2015;64(6):2207–19. <https://doi.org/10.2337/db14-1011>.
134. Saltiel AR, Kahn CR. Insulin signalling and the regulation of glucose and lipid metabolism. *Nature (London)*. 2001;414(6865):799–806. <https://doi.org/10.1038/414799a>.
135. Hofbauer SW, et al. The AKT1 isoform plays a dominant role in the survival and chemoresistance of chronic lymphocytic leukaemia cells. *Br J Haematol*. 2016;172(5):815–9. <https://doi.org/10.1111/bjh.13542>.
136. Yu H, Littlewood T, Bennett M. Akt isoforms in vascular disease. *Vascular Pharmacol*. 2015;71:57–64. <https://doi.org/10.1016/j.vph.2015.03.003>.
137. George S, et al. A family with severe insulin resistance and diabetes due to a mutation in AKT2. *Science*. 2004;304(5675):1325–8. <https://doi.org/10.1126/science.1096706>.
138. Sun W, et al. The beneficial effects of Zn on Akt-mediated insulin and cell survival signaling pathways in diabetes. *J Trace Elem Med Biol*. 2018;46:117–27. <https://doi.org/10.1016/j.jtemb.2017.12.005>.
139. Sanchez-Garrido J, Shenoy AR. Regulation and repurposing of nutrient sensing and autophagy in innate immunity. *Autophagy*. 2020. <https://doi.org/10.1080/15548627.2020.1783119>.
140. Yamauchi T, et al. The fat-derived hormone adiponectin reverses insulin resistance associated with both lipotrophy and obesity. *Nat Med*. 2001;7(8):941–6. <https://doi.org/10.1038/90984>.
141. Chen H, et al. Adiponectin activates the AMPK signaling pathway to regulate lipid metabolism in bovine hepatocytes. *J Steroid Biochem Mol Biol*. 2013;138:445–54. <https://doi.org/10.1016/j.jsbmb.2013.08.013>.
142. Leonardini A, et al. Cross-talk between PPARgamma and insulin signaling and modulation of insulin sensitivity. *PPAR Res*. 2009;2009:818945.
143. Nawrocki AR, et al. Mice lacking adiponectin show decreased hepatic insulin sensitivity and reduced responsiveness to peroxisome proliferator-activated receptor gamma agonists. *J Biol Chem*. 2006;281(5):2654–60. <https://doi.org/10.1074/jbc.M505311200>.
144. Tomas E, et al. Enhanced muscle fat oxidation and glucose transport by ACRP30 globular domain: acetyl-CoA carboxylase inhibition and AMP-activated protein kinase activation. *Proc Natl Acad Sci U S A*. 2002;99(25):16309–13. <https://doi.org/10.1073/pnas.222657499>.
145. Ceddia RB, et al. Globular adiponectin increases GLUT4 translocation and glucose uptake but reduces glycogen synthesis in rat skeletal muscle cells. *Diabetologia*. 2005;48(1):132–9. <https://doi.org/10.1007/s00125-004-1609-y>.
146. Jung UJ, Choi MS. Obesity and its metabolic complications: the role of adipokines and the relationship between obesity, inflammation, insulin resistance, dyslipidemia and nonalcoholic fatty liver disease. *Int J Mol Sci*. 2014;15(4):6184–223. <https://doi.org/10.3390/ijms15046184>.
147. Liu M, et al. Fat-specific DsbA-L overexpression promotes adiponectin multimerization and protects mice from diet-induced obesity and insulin resistance. *Diabetes*. 2012;61(11):2776–86. <https://doi.org/10.2337/db12-0169>.
148. Zhou X, Li J, Wei L. Silencing of DsbA-L gene impairs the PPAR γ agonist function of improving insulin resistance in a high-glucose cell model. *J Zhejiang Univ Sci B*. 2020;21(12):990–8. <https://doi.org/10.1631/jzus.B2000432>.

149. Wang Y, et al. Cyclic GMP-AMP synthase is required for cell proliferation and inflammatory responses in rheumatoid arthritis synoviocytes. *Mediators Inflamm.* 2015;2015: 192329. <https://doi.org/10.1155/2015/192329>.
150. Kaur S, et al. Role of the Akt pathway in mRNA translation of interferon-stimulated genes. *Proc Natl Acad Sci U S A.* 2008;105(12):4808–13. <https://doi.org/10.1073/pnas.0710907105>.
151. Kaur S, et al. Dual regulatory roles of phosphatidylinositol 3-kinase in IFN signaling. *J Immunol.* 2008;181(10):7316–23. <https://doi.org/10.4049/jimmunol.181.10.7316>.
152. Bodur C, et al. The IKK-related kinase TBK1 activates mTORC1 directly in response to growth factors and innate immune agonists. *EMBO J.* 2018;37(1):19–38. <https://doi.org/10.15252/emboj.201696164>.
153. Hasan M, et al. Chronic innate immune activation of TBK1 suppresses mTORC1 activity and dysregulates cellular metabolism. *Proc Natl Acad Sci.* 2017;114(4):746–51. <https://doi.org/10.1073/pnas.1611131114>.
154. Gong Y, et al. Double knockout of Akt2 and AMPK accentuates high fat diet-induced cardiac anomalies through a cGAS–STING-mediated mechanism. *Biochim Biophys Acta (BBA) Mol Basis Dis.* 2020;1866(10): 165855. <https://doi.org/10.1016/j.bbadis.2020.165855>.
155. Shirakawa K, et al. Obesity accelerates T cell senescence in murine visceral adipose tissue. *J Clin Invest.* 2016;126(12):4626–39. <https://doi.org/10.1172/JCI88606>.
156. Xia C, Rao X, Zhong J. Role of T lymphocytes in type 2 diabetes and diabetes-associated inflammation. *J Diabetes Res.* 2017;2017:6494795. <https://doi.org/10.1155/2017/6494795>.
157. Roep BO. The role of T-cells in the pathogenesis of Type 1 diabetes: from cause to cure. *Diabetologia.* 2003;46(3):305–21. <https://doi.org/10.1007/s00125-003-1089-5>.
158. Tan Y, et al. Mechanisms of diabetic cardiomyopathy and potential therapeutic strategies: preclinical and clinical evidence. *Nat Rev Cardiol.* 2020;17(9):585–607. <https://doi.org/10.1038/s41569-020-0339-2>.
159. Ge T, et al. The adaptive immune role of metallothioneins in the pathogenesis of diabetic cardiomyopathy: good or bad. *Am J Physiol Heart Circ Physiol.* 2019;317(2):H264–75. <https://doi.org/10.1152/ajpheart.00123.2019>.
160. Larkin B, et al. Cutting edge: activation of STING in T cells induces type I IFN responses and cell death. *J Immunol.* 2017;199(2):397–402. <https://doi.org/10.4049/jimmunol.1601999>.
161. Cerboni S, et al. Intrinsic antiproliferative activity of the innate sensor STING in T lymphocytes. *J Exp Med.* 2017;214(6):1769–85. <https://doi.org/10.1084/jem.20161674>.
162. Deng L, et al. STING-dependent cytosolic DNA sensing promotes radiation-induced type I interferon-dependent antitumor immunity in immunogenic tumors. *Immunity.* 2014;41(5):843–52. <https://doi.org/10.1016/j.immuni.2014.10.019>.
163. Demaria O, et al. STING activation of tumor endothelial cells initiates spontaneous and therapeutic antitumor immunity. *Proc Natl Acad Sci.* 2015;112(50):15408–13. <https://doi.org/10.1073/pnas.1512832112>.
164. Wang H, et al. cGAS is essential for the antitumor effect of immune checkpoint blockade. *Proc Natl Acad Sci U S A.* 2017;114(7):1637–42. <https://doi.org/10.1073/pnas.1621363114>.
165. Wu J, et al. STING-mediated disruption of calcium homeostasis chronically activates ER stress and primes T cell death. *J Exp Med.* 2019;216(4):867–83. <https://doi.org/10.1084/jem.20182192>.
166. Imanishi T, Saito T. T cell co-stimulation and functional modulation by innate signals. *Trends Immunol.* 2020;41(3):200–12. <https://doi.org/10.1016/j.it.2020.01.003>.
167. Zhou H, et al. DsbA-L deficiency in T cells promotes diet-induced thermogenesis through suppressing IFN-gamma production. *Nat Commun.* 2021;12(1):326. <https://doi.org/10.1038/s41467-020-20665-4>.
168. Dinh W, et al. Elevated plasma levels of TNF-alpha and interleukin-6 in patients with diastolic dysfunction and glucose metabolism disorders. *Cardiovasc Diabetol.* 2009;8:58. <https://doi.org/10.1186/1475-2840-8-58>.
169. Masters SL, Latz E, O'Neill LA. The inflammasome in atherosclerosis and type 2 diabetes. *Sci Transl Med.* 2011; 3(81): 81ps17. <https://doi.org/10.1126/scitranslmed.3001902>.
170. Bajpai A, Tilley DG. The role of leukocytes in diabetic cardiomyopathy. *Front Physiol.* 2018;9:1547. <https://doi.org/10.3389/fphys.2018.01547>.
171. Gordon JW, Shaw JA, Kirshenbaum LA. Multiple facets of NF-kappaB in the heart: to be or not to NF-kappaB. *Circ Res.* 2011;108(9):1122–32. <https://doi.org/10.1161/CIRCRESAHA.110.226928>.
172. Wende AR, et al. Glucose transporter 4-deficient hearts develop maladaptive hypertrophy in response to physiological or pathological stresses. *Am J Physiol Heart Circ Physiol.* 2017;313(6):H1098–108. <https://doi.org/10.1152/ajpheart.00101.2017>.
173. Cho H, et al. Insulin resistance and a diabetes mellitus-like syndrome in mice lacking the protein kinase Akt2 (PKB beta). *Science.* 2001;292(5522):1728–31. <https://doi.org/10.1126/science.292.5522.1728>.
174. Garofalo RS, et al. Severe diabetes, age-dependent loss of adipose tissue, and mild growth deficiency in mice lacking Akt2/PKB beta. *J Clin Invest.* 2003;112(2):197–208. <https://doi.org/10.1172/JCI16885>.
175. Guo Y, et al. Mitochondrial DNA drives noncanonical inflammation activation via cGAS–STING signaling pathway in retinal microvascular endothelial cells. *Cell Commun Signal.* 2020;18(1):172. <https://doi.org/10.1186/s12964-020-00637-3>.
176. Hu J, et al. Exosomal Mst1 transfer from cardiac microvascular endothelial cells to cardiomyocytes deteriorates diabetic cardiomyopathy. *Biochim Biophys Acta Mol Basis Dis.* 2018;1864(11):3639–49. <https://doi.org/10.1016/j.bbadis.2018.08.026>.
177. Teng X, et al. Selective deletion of endothelial cell calpain in mice reduces diabetic cardiomyopathy by improving angiogenesis. *Diabetologia.* 2019;62(5):860–72. <https://doi.org/10.1007/s00125-019-4828-y>.
178. Shen X, et al. Cardiac mitochondrial damage and biogenesis in a chronic model of type 1 diabetes. *Am J Physiol Endocrinol Metab.* 2004;287(5):E896–905. <https://doi.org/10.1152/ajpendo.00047.2004>.
179. Cividini F, et al. O-GlcNAcylation of 8-oxoguanine DNA Glycosylase (Ogg1) impairs oxidative mitochondrial DNA lesion repair in diabetic hearts. *J Biol Chem.* 2016;291(51):26515–28. <https://doi.org/10.1074/jbc.M116.754481>.
180. Jiang S, et al. Diabetic-induced alterations in hepatic glucose and lipid metabolism: the role of type 1 and type 2 diabetes mellitus (Review). *Mol Med Rep.* 2020;22(2):603–11. <https://doi.org/10.3892/mmr.2020.11175>.
181. Cho CS, et al. Lipotoxicity induces hepatic protein inclusions through TANK binding kinase 1-mediated p62/sequestosome 1 phosphorylation. *Hepatology.* 2018;68(4):1331–46. <https://doi.org/10.1002/hep.29742>.
182. Anders HJ, et al. CKD in diabetes: diabetic kidney disease versus nondiabetic kidney disease. *Nat Rev Nephrol.* 2018;14(6):361–77. <https://doi.org/10.1038/s41581-018-0001-y>.
183. Schena FP, Gesualdo L. Pathogenetic mechanisms of diabetic nephropathy. *J Am Soc Nephrol.* 2005;16(Suppl 1):S30–3. <https://doi.org/10.1681/asn.2004110970>.
184. Thomas MC, et al. Diabetic kidney disease. *Nat Rev Dis Prim.* 2015. <https://doi.org/10.1038/nrdp.2015.18>.
185. Berthier CC, et al. Enhanced expression of Janus kinase-signal transducer and activator of transcription pathway members in human diabetic nephropathy. *Diabetes.* 2009;58(2):469–77. <https://doi.org/10.2337/db08-1328>.
186. Starkey JM, et al. Diabetes-induced activation of canonical and noncanonical nuclear factor- κ B pathways in renal cortex. *Diabetes.* 2006;55(5):1252–9. <https://doi.org/10.3892/mmr.2020.11175>.
187. Porrini E, et al. Non-proteinuric pathways in loss of renal function in patients with type 2 diabetes. *Lancet Diabetes Endocrinol.* 2015;3(5):382–91. [https://doi.org/10.1016/S2213-8587\(15\)00094-7](https://doi.org/10.1016/S2213-8587(15)00094-7).
188. Luis-Rodríguez D. Pathophysiological role and therapeutic implications of inflammation in diabetic nephropathy. *World J Diabetes.* 2012;3(1):7. <https://doi.org/10.4239/wjdv.v3.i1.7>.
189. Forbes JM, Thorburn DR. Mitochondrial dysfunction in diabetic kidney disease. *Nat Rev Nephrol.* 2018;14(5):291–312. <https://doi.org/10.1038/nrneph.2018.9>.
190. Sallstrom J, et al. Inhibition of sodium-linked glucose reabsorption normalizes diabetes-induced glomerular hyperfiltration in conscious adenosine A(1)-receptor deficient mice. *Acta Physiol (Oxf).* 2014;210(2):440–5. <https://doi.org/10.1111/apha.12152>.
191. Oyarzun C, et al. Increased levels of adenosine and ecto 5'-nucleotidase (CD73) activity precede renal alterations in experimental diabetic rats.

Biochem Biophys Res Commun. 2015;468(1–2):354–9. <https://doi.org/10.1016/j.bbrc.2015.10.095>.

192. Kretschmar C, et al. Reduced adenosine uptake and its contribution to signaling that mediates profibrotic activation in renal tubular epithelial cells: implication in diabetic nephropathy. PLoS ONE. 2016;11(1): e0147430. <https://doi.org/10.1371/journal.pone.0147430>.
193. Zhang Y, et al. Streptavidin promotes DNA binding and activation of cGAS to enhance innate immunity. iScience. 2020;23(9): 101463. <https://doi.org/10.1016/j.isci.2020.101463>.
194. Wang Y, et al. Protocol for monitoring DNA-triggered cGAS/STING signaling in mammalian cells and mice. STAR Protoc. 2020;1(3): 100171. <https://doi.org/10.1016/j.xpro.2020.100171>.

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